412 Rec'd PCT/PTO 24 MAY 2000

FORM PTO-1390 (Modified) (REV 11-98) U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371

ATTORNEY'S DOCKET NUMBER: 13627

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.5): 09/555108

INTERNATIONAL APPLICATION NO.	INTERNATIONAL FILING DATE	PRIORITY DATE CLAIMED
PCT/AU98/00981	26 November 1998 (26.11.98)	26 November 1997 (26.11.97)

TITLE OF INVENTION
COMPOSITIONS OF LEUKAEMIA INHIBITORY FACTOR

APPLICANT(S) FOR DO/EO/US
Susan Ann Charman and Anthony John Radford

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
   a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
   b. ☒ has been transmitted by the International Bureau.
   c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ A copy of the International Search Report (PCT/ISA/210).
8. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
   a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
   b. ☐ have been transmitted by the International Bureau.
   c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
   d. ☒ have not been made and will not be made.
9. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
10. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).

Items 13 to 20 below concern document(s) or information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☒ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☒ Certificate of Mailing by Express Mail
20. ☒ Other items or information:

Courtsey copy of international application
Five (5) sheets of drawings

PCTUS1/REV03

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.5)	INTERNATIONAL APPLICATION NO.	ATTORNEY'S DOCKET NUMBER
09/555108	PCT/AU98/00981	13627

21. The following fees are submitted:.

BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :

		CALCULATIONS PTO USE ONLY
☒	Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2) paid to USPTO and International Search Report not prepared by the EPO or JPO	**$970.00**
☐	International preliminary examination fee (37 CFR 1.482) not paid to USPTO but Internation Search Report prepared by the EPO or JPO	**$840.00**
☐	International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO	**$690.00**
☐	International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4)	**$670.00**
☐	International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4)	**$96.00**

ENTER APPROPRIATE BASIC FEE AMOUNT =	**$970.00**	
Surcharge of **$130.00** for furnishing the oath or declaration later than ☐ 20 ☒ 30 months from the earliest claimed priority date (37 CFR 1.492 (e)).	**$130.00**	

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	43 - 20 =	23	x $18.00	**$414.00**	
Independent claims	4 - 3 =	1	x $78.00	**$78.00**	
Multiple Dependent Claims **(check if applicable)**.			☐	**$0.00**	
TOTAL OF ABOVE CALCULATIONS =				**$1,592.00**	
Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28) **(check if applicable)**.			☐	**$0.00**	
SUBTOTAL =				**$1,592.00**	
Processing fee of **$130.00** for furnishing the English translation later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492 (f)). +				**$0.00**	
TOTAL NATIONAL FEE =				**$1,592.00**	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) **(check if applicable).**			☐	**$0.00**	
TOTAL FEES ENCLOSED =				**$1,592.00**	
				Amount to be: refunded	$
				charged	$

☒ A check in the amount of **$1,592.00** to cover the above fees is enclosed.

☐ Please charge my Deposit Account No. in the amount of to cover the above fees.
A duplicate copy of this sheet is enclosed.

☒ The Commissioner is hereby authorized to charge any fees which may be required, or credit any overpayment to Deposit Account No. **19-1013/SSMP** A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

Leopold Presser
Registration No. 19, 827

Scully, Scott, Murphy & Presser
400 Garden City Plaza
Garden City, NY 11530
(516)742-4343

SIGNATURE

Leopold Presser
NAME

19,827
REGISTRATION NUMBER

May 24, 2000
DATE

PATENTS

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s): Susan Ann Charman & Anthony John Radford

Examiner: Unassigned

Serial No.: To be assigned

Art Unit: Unassigned

Filed: Herewith

Docket: 13627

Int'l Appln. No. PCT/AU98/00981

Dated: May 24, 2000

Int'l Filing Date: November 26, 1998

For: COMPOSITIONS OF LEUKAEMIA INHIBITORY FACTOR

Assistant Commissioner for Patents
Washington, DC 20231

PRELIMINARY AMENDMENT

Sir:

In connection with the filing of the above-identified application, kindly enter the following amendments:

IN THE CLAIMS:

Please amend claims 3, 4, 6, 16, 19, 20, 30, 32, 41 and 42 as follows:

Claim 3, line 1, delete "or 2";

Claim 4, line 1, delete "or 2 or 3";

CERTIFICATE OF MAILING BY "EXPRESS MAIL"

"Express Mail" Mailing Label Number: EL308568422US
Date of Deposit: May 24, 2000

I hereby certify that this correspondence is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 C.F.R. §1.10 on the date indicated above and is addressed to the Assistant Commissioner of Patents and Trademarks, Washington, D.C. 20231

Dated: May 24, 2000

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Claim 6, line 1, delete "or 5";

Claim 16, line 1, delete "or 7";

Claim 19, line 1, delete " 17 or 18";

Claim 20, line 1, delete "or 14";

Claim 30, line 1, amend "and of claims 22 to 29" to read --claim 21--;

Claim 32, line 1, amend "any one of the claims 22 to 31" to read --claim 21--;

Claim 41, line 1, amend "any one of claims 33 to 40" to read --claim 33--; and

Claim 42, line 1, amend "any one of claims 41" to read --claim 41--.

Please add claim 43 as follows:

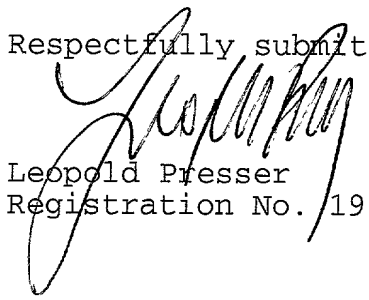
--43. A composition according to claim 14 wherein LIF is present in an amount from about 0.1 µg/ml to about 100 mg/ml.--

REMARKS

As originally prepared, several claims did not comply with the multiple dependent claim style specified by U.S. law. The amendments submitted herewith have been made to delete all multiple dependancies. The amendments submitted above do not introduce any new subject matter.

It is respectfully requested that the above amendments be entered before an action on the merits is issued.

Respectfully submitted,



Leopold Presser
Registration No. 19,827

SCULLY, SCOTT, MURPHY & PRESSER
400 Garden City Plaza
Garden City, New York 11530
(516) 742-4343

LP/am

COMPOSITIONS OF LEUKAEMIA INHIBITORY FACTOR

FIELD OF THE INVENTION

The present invention relates generally to compositions and more particularly to compositions comprising leukaemia inhibitory factor (hereinafter referred to as "LIF") or derivative or homologues thereof. The compositions of the present invention are particularly useful as compositions which exhibit enhanced stability and/or which exhibit reduced aggregation and/or reduced deamidation of LIF, its derivatives or other active ingredients .

BACKGROUND OF THE INVENTION

LIF is a polyfunctional glycoprotein with diverse actions on a broad range of tissue and cell types, including induction of differentiation in a number of myeloid leukaemic cell lines, suppression of differentiation in normal embryonic stem cells, stimulation of proliferation of osteoblasts and DA-1 haemopoietic cells and potentiation of the of the proliferative action of interleukin-3 (IL-3) on megakaryocyte precursors. Functionally, LIF is able to switch autonomic nerve signalling from adrenergic to cholinergic mode, stimulate calcium release from bones, stimulate the production of acute phase proteins by hepatocytes and induce loss of fat deposits by inhibiting lipoprotein lipase-mediated lipid transport into adipocytes.

With a potentially broad range of clinical applications, it is imperative that compositions containing LIF are presented in a stable form and remain so during an extended period which may include shipment, handling and storage. Thus, a stable composition is one which retains its physical, chemical, therapeutic and toxicological profile over this period.

Deamidation is the most significant chemical degradation of LIF over time. It is clearly desirable that this process is minimized. Physical degradation, such as aggregation or flocculation, may occur due to denaturation caused by elevated temperatures and/or agitation and excessive handling of the composition. Such degradation is clearly undesirable in terms of appearance and more importantly, consistent and effective administration of LIF in clinical applications. Storage

at temperatures below room temperature typically retards chemical degradation, with storage in the frozen state being generally the most effective. Whilst this may minimize chemical degradation, the process of thawing the composition may then result in aggregation.

Thus, there exists a need for a stable composition and, in particular, a stable pharmaceutical composition of LIF and/or its derivatives or homologues wherein chemical and physical degradation is minimised.

SUMMARY OF THE INVENTION

Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated integer or group of integers but not the exclusion of any other integer or group of integers.

One aspect of the present invention contemplates a composition comprising leukaemia inhibitory factor (LIF) or a derivative or homologue thereof and a stabilizing agent facilitating chemical and/or physical stability of LIF in the composition and one or more pharmaceutically acceptable carriers and/or diluents.

Another aspect of the invention provides a composition with improved chemical and physical stability comprising LIF or a derivative or homologue thereof, a stabilizing agent, and one or more pharmaceutically acceptable carriers or diluents under conditions in which aggregation of LIF is reduced.

Yet another aspect of the invention provides a composition with improved chemical and physical stability comprising LIF or a derivative or homologue thereof, a stabilizing agent, and one or more pharmaceutically acceptable carriers or diluents under conditions in which deamidation of LIF is reduced.

Still another aspect the present invention is directed to a stable composition comprising LIF or a derivative or homologue thereof, together with one or more pharmaceutically acceptable carriers or diluents, wherein the composition has a pH of between about 3.5 and about 6.5.

A further aspect the present invention provides a stable composition comprising LIF or a derivative or homologue thereof, together with one or more pharmaceutically acceptable carriers or diluents, wherein the composition has a pH of between about 3.5 and about 6.5 under conditions in which aggregation of LIF is reduced.

Another aspect the present invention contemplates a stable composition comprising LIF or a derivative or homologue thereof, together with one or more pharmaceutically acceptable carriers or diluents, wherein the composition has a pH of between about 3.5 and about 6.5 under conditions in which deamidation of LIF is reduced.

Yet another aspect of the present invention contemplates a method for preparing a composition comprising Leukaemia Inhibition Factor (LIF) or a derivative or homologue thereof and which exhibits reduced deamidation and/or agglutination or LIF or a derivative or homologue over time said method comprising admixing LIF or its derivative or homologue with a stabilizing agent.

Still another aspect of the present invention is directed to the use of a stabilizing agent in the manufacture of a composition exhibiting improved chemical and/or physical stability of Leukaemic Inhibitory Factor (LIF) or a derivative or homologue thereof.

Preferred compositions in accordance with the present invention are referred to as "pharmaceutical compositions" where LIF or its derivatives or homologues is/are present in a pharmaceutically acceptable composition.

BRIEF DESCRIPTION OF THE FIGURES

Figures 1 to 3, respectively, are a diagrammatic representations of Reversed Phase, Ion Exchange and Size Exclusion chromatograms for a 1.0 mg/ml standard solution of LIF prepared as described in Example 1 by diluting "stock" solution with 2 mM phosphate buffer, pH 6.42, containing 0.01% polysorbate.

Figure 4 is a graphical representation showing LIF concentration for samples at each pH after freeze/thaw cycling.

Figure 5 is a graphical representation of the average concentration over 5 freeze/thaw cycles for each pH value.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention provides compositions comprising LIF or its derivatives or homologues. The present invention particularly provides LIF or related molecules in a stable form.

Unless otherwise specified, the term "LIF" or "Leukaemia Inhibitory Factor" refers herein to synthetic, recombinant or purified naturally occurring LIF from animal or avian species. Preferred animal species are mammals such as humans, primates and livestock animals as well as any or all derivatives or homologues of LIF (e.g. sheep, pigs, cows, goats, donkeys and horses), laboratory animals (e.g. murine species, guinea pigs, rabbits and hamsters), companion animals (e.g. dogs and cats) or captive wild animals (e.g. kangaroos, foxes, and deer). Preferred avian species include but are not limited to caged birds, chickens, ducks, geese and game birds. As referred to here, LIF or Leukaemia Inhibitory Factor includes reference to derivatives, homologues and analogues of LIF. Derivatives, homologues, mimetics and analogues include parts, fragments or portions of LIF which are functionally active or which otherwise have a useful biological activity (eg. as an antagonist, antigen to induce antibody formation, as a diagnostic agent or as a therapeutic molecule). Such derivatives or parts thereof include any one or more contiguous series of amino acids contained within any one of the above LIF molecules

and includes single or multiple amino acids substitutions, deletions and/or additions to or in the natural, synthetic or recombinant LIF molecule as well as hyperglycosolated and deglycosolated forms. Conditions for preparing recombinant LIF are disclosed in International Patent Application Nos PCT/AU88/00093 and PCT/AU90/00001 although these conditions may vary depending on the host cell used. Any such variations and/or modifications are within the scope of the subject invention. The host cells may be eukaryotic (eg. yeast, mammalian, insect, plant etc) or prokaryotic (eg. *Escherichia coli*, *Bacillus* sp, *Pseudomonas* sp etc) cells.

Analogues and mimetics include molecules which contain non-naturally occurring amino acids or which do not contain amino acids but nevertheless behave functionally the same as or similar to LIF. Natural product screening is one useful strategy for identifying analogues and mimetics. Analogues of LIF contemplated herein also include modifications to side chains, incorporation of unnatural amino acids and/or their derivatives during peptide synthesis and the use of cross linkers and other methods which impose conformational constraints on the protein molecule or their analogues.

Examples of side chain modifications contemplated by the present invention include modifications of amino groups such as by reductive alkylation by reaction with an aldehyde followed by reduction with $NaBH_4$; amidination with methylacetimidate; acylation with acetic anhydride; carbamoylation of amino groups with cyanate; trinitrobenzylation of amino groups with 2, 4, 6-trinitrobenzene sulphonic acid (TNBS); acylation of amino groups with succinic anhydride and tetrahydrophthalic anhydride; and pyridoxylation of lysine with pyridoxal-5-phosphate followed by reduction with $NaBH_4$.

The guanidine group of arginine residues may be modified by the formation of heterocyclic condensation products with reagents such as 2,3-butanedione, phenylglyoxal and glyoxal.

The carboxyl group may be modified by carbodiimide activation *via* O-acylisourea formation followed by subsequent derivitisation, for example, to a corresponding amide.

Sulphydryl groups may be modified by methods such as carboxymethylation with iodoacetic acid

or iodoacetamide; performic acid oxidation to cysteic acid; formation of a mixed disulphides with other thiol compounds; reaction with maleimide, maleic anhydride or other substituted maleimide; formation of mercurial derivatives using 4-chloromercuribenzoate, 4-chloromercuriphenylsulphonic acid, phenylmercury chloride, 2-chloromercuri-4-nitrophenol and other mercurials; carbamoylation with cyanate at alkaline pH.

Tryptophan residues may be modified by, for example, oxidation with N-bromosuccinimide or alkylation of the indole ring with 2-hydroxy-5-nitrobenzyl bromide or sulphenyl halides. Tyrosine residues on the other hand, may be altered by nitration with tetranitromethane to form a 3-nitrotyrosine derivative.

Modification of the imidazole ring of a histidine residue may be accomplished by alkylation with iodoacetic acid derivatives or N-carboethoxylation with diethylpyrocarbonate.

Examples of incorporating unnatural amino acids and derivatives during protein synthesis include, but are not limited to, use of norleucine, 4-amino butyric acid, 4-amino-3-hydroxy-5-phenylpentanoic acid, 6-aminohexanoic acid, t-butylglycine, norvaline, phenylglycine, ornithine, sarcosine, 4-amino-3-hydroxy-6-methylheptanoic acid, 2-thienyl alanine and/or D-isomers of amino acids. A list of unnatural amino acid contemplated herein is shown in Table 1.

TABLE 1

Non-conventional amino acid	Code	Non-conventional amino acid	Code
α-aminobutyric acid	Abu	L-N-methylalanine	Nmala
α-amino-α-methylbutyrate	Mgabu	L-N-methylarginine	Nmarg
aminocyclopropane-carboxylate	Cpro	L-N-methylasparagine	Nmasn
		L-N-methylaspartic acid	Nmasp
aminoisobutyric acid	Aib	L-N-methylcysteine	Nmcys
aminonorbornyl-carboxylate	Norb	L-N-methylglutamine	Nmgln
		L-N-methylglutamic acid	Nmglu
cyclohexylalanine		Chexa L-N-methylhistidine	Nmhis
cyclopentylalanine	Cpen	L-N-methylisolleucine	Nmile
D-alanine	Dal	L-N-methylleucine	Nmleu
D-arginine	Darg	L-N-methyllysine	Nmlys
D-aspartic acid	Dasp	L-N-methylmethionine	Nmmet
D-cysteine	Dcys	L-N-methylnorleucine	Nmnle
D-glutamine	Dgln	L-N-methylnorvaline	Nmnva
D-glutamic acid	Dglu	L-N-methylornithine	Nmorn
D-histidine	Dhis	L-N-methylphenylalanine	Nmphe
D-isoleucine	Dile	L-N-methylproline	Nmpro
D-leucine	Dleu	L-N-methylserine	Nmser
D-lysine	Dlys	L-N-methylthreonine	Nmthr
D-methionine	Dmet	L-N-methyltryptophan	Nmtrp
D-ornithine	Dorn	L-N-methyltyrosine	Nmtyr
D-phenylalanine	Dphe	L-N-methylvaline	Nmval
D-proline	Dpro	L-N-methylethylglycine	Nmetg
D-serine	Dser	L-N-methyl-t-butylglycine	Nmtbug
D-threonine	Dthr	L-norleucine	Nle
D-tryptophan	Dtrp	L-norvaline	Nva
D-tyrosine	Dtyr	α-methyl-aminoisobutyrate	Maib

D-valine	Dval	α-methyl-γ-aminobutyrate	Mgabu
D-α-methylalanine	Dmala	α-methylcyclohexylalanine	Mchexa
D-α-methylarginine	Dmarg	α-methylcylcopentylalanine	Mcpen
D-α-methylasparagine	Dmasn	α-methyl-α-napthylalanine	Manap
D-α-methylaspartate	Dmasp	α-methylpenicillamine	Mpen
D-α-methylcysteine	Dmcys	N-(4-aminobutyl)glycine	Nglu
D-α-methylglutamine	Dmgln	N-(2-aminoethyl)glycine	Naeg
D-α-methylhistidine	Dmhis	N-(3-aminopropyl)glycine	Norn
D-α-methylisoleucine	Dmile	N-amino-α-methylbutyrate	Nmaabu
D-α-methylleucine	Dmleu	α-napthylalanine	Anap
D-α-methyllysine	Dmlys	N-benzylglycine	Nphe
D-α-methylmethionine	Dmmet	N-(2-carbamylethyl)glycine	Ngln
D-α-methylornithine	Dmorn	N-(carbamylmethyl)glycine	Nasn
D-α-methylphenylalanine	Dmphe	N-(2-carboxyethyl)glycine	Nglu
D-α-methylproline	Dmpro	N-(carboxymethyl)glycine	Nasp
D-α-methylserine	Dmser	N-cyclobutylglycine	Ncbut
D-α-methylthreonine	Dmthr	N-cycloheptylglycine	Nchep
D-α-methyltryptophan	Dmtrp	N-cyclohexylglycine	Nchex
D-α-methyltyrosine	Dmty	N-cyclodecylglycine	Ncdec
D-α-methylvaline	Dmval	N-cylcododecylglycine	Ncdod
D-N-methylalanine	Dnmala	N-cyclooctylglycine	Ncoct
D-N-methylarginine	Dnmarg	N-cyclopropylglycine	Ncpro
D-N-methylasparagine	Dnmasn	N-cycloundecylglycine	Ncund
D-N-methylaspartate	Dnmasp	N-(2,2-diphenylethyl)glycine	Nbhm
D-N-methylcysteine	Dnmcys	N-(3,3-diphenylpropyl)glycine	Nbhe
D-N-methylglutamine	Dnmgln	N-(3-guanidinopropyl)glycine	Narg
D-N-methylglutamate	Dnmglu	N-(1-hydroxyethyl)glycine	Nthr
D-N-methylhistidine	Dnmhis	N-(hydroxyethyl))glycine	Nser
D-N-methylisoleucine	Dnmile	N-(imidazolylethyl))glycine	Nhis
D-N-methylleucine	Dnmleu	N-(3-indolylyethyl)glycine	Nhtrp
D-N-methyllysine	Dnmlys	N-methyl-γ-aminobutyrate	Nmgabu

N-methylcyclohexylalanine	Nmchexa	D-N-methylmethionine	Dnmmet
D-N-methylornithine	Dnmorn	N-methylcyclopentylalanine	Nmcpen
N-methylglycine	Nala	D-N-methylphenylalanine	Dnmphe
N-methylaminoisobutyrate	Nmaib	D-N-methylproline	Dnmpro
N-(1-methylpropyl)glycine	Nile	D-N-methylserine	Dnmser
N-(2-methylpropyl)glycine	Nleu	D-N-methylthreonine	Dnmthr
D-N-methyltryptophan	Dnmtrp	N-(1-methylethyl)glycine	Nval
D-N-methyltyrosine	Dnmtyr	N-methyla-napthylalanine	Nmanap
D-N-methylvaline	Dnmval	N-methylpenicillamine	Nmpen
γ-aminobutyric acid	Gabu	N-(*p*-hydroxyphenyl)glycine	Nhtyr
L-*t*-butylglycine	Tbug	N-(thiomethyl)glycine	Ncys
L-ethylglycine	Etg	penicillamine	Pen
L-homophenylalanine	Hphe	L-α-methylalanine	Mala
L-α-methylarginine	Marg	L-α-methylasparagine	Masn
L-α-methylaspartate	Masp	L-α-methyl-*t*-butylglycine	Mtbug
L-α-methylcysteine	Mcys	L-methylethylglycine	Metg
L-α-methylglutamine	Mgln	L-α-methylglutamate	Mglu
L-α-methylhistidine	Mhis	L-α-methylhomophenylalanine	Mhphe
L-α-methylisoleucine	Mile	N-(2-methylthioethyl)glycine	Nmet
L-α-methylleucine	Mleu	L-α-methyllysine	Mlys
L-α-methylmethionine	Mmet	L-α-methylnorleucine	Mnle
L-α-methylnorvaline	Mnva	L-α-methylornithine	Morn
L-α-methylphenylalanine	Mphe	L-α-methylproline	Mpro
L-α-methylserine	Mser	L-α-methylthreonine	Mthr
L-α-methyltryptophan	Mtrp	L-α-methyltyrosine	Mtyr
L-α-methylvaline	Mval	L-N-methylhomophenylalanine	Nmhphe

N-(N-(2,2-diphenylethyl) carbamylmethyl)glycine	Nnbhm	N-(N-(3,3-diphenylpropyl) carbamylmethyl)glycine	Nnbhe
1-carboxy-1-(2,2-diphenyl-ethylamino)cyclopropane	Nmbc		

Crosslinkers can be used, for example, to stabilise 3D conformations, using homo-bifunctional crosslinkers such as the bifunctional imido esters having $(CH_2)_n$ spacer groups with n=1 to n=6, glutaraldehyde, N-hydroxysuccinimide esters and hetero-bifunctional reagents which usually contain an amino-reactive moiety such as N-hydroxysuccinimide and another group specific-reactive moiety such as maleimido or dithio moiety (SH) or carbodiimide (COOH). In addition, peptides can be conformationally constrained by, for example, incorporation of C_α and N_α-methylamino acids, introduction of double bonds between C_α and C_β atoms of amino acids and the formation of cyclic peptides or analogues by introducing covalent bonds such as forming an amide bond between the N and C termini, between two side chains or between a side chain and the N or C terminus.

All these types of modifications may be important to further stabilise LIF in the composition of the present invention.

The compositions of the present invention achieve their stability through judicious choice of pH conditions within the range of between from about 3.5 to about 6.5 inclusive and optionally the presence of one or more suitable stabilizing agents. Preferably, the pH range is between from about 4.0 - 6.0 inclusive, more preferably between from about 4.5 to about 5.5 inclusive. Most preferably, the pH of the composition is about 5.0.

Accordingly, another aspect of the present invention provides a composition comprising Leukaemia Inhibitory Factor (LIF) and one or more pharmaceutically acceptable carriers and/or diluents and wherein the composition has a pH of between about 3.5 and 6.5.

Suitable stabilizing agents are known to those skilled in the art and include isotonicity agents,

agents to increase or maintain the conformational stability of LIF and surfactants. It is understood that one agent may possess more than one stabilizing property and more than one agent may be employed to achieve a stabilizing effect.

Suitable isotonicity agents are those which maintain approximately the same osmotic pressure as that of cellular fluids, and are known to those skilled in the art. These may include, but are not limited to, polyhydric alcohols such as sorbitol, pharmaceutically acceptable salts such as NaCl, buffer species, sugars and pharmaceutically acceptable polymeric compounds. Suitable surfactants may be anionic, cationic, amphoteric or non-ionic. Preferred surfactants include fatty alcohols such as lauryl, cetyl and stearyl alcohols, glyceryl esters such as the mono-, di- and triglycerides, fatty acid esters of fatty alcohols and other alcohols such as propylene glycol, polyethylene glycol, sorbitol, sucrose and cholesterol. Other suitable agents include the polysorbates such as polysorbates 20, 40, 60 and 80 and sorbitan ester, polyoxyethylene derivatives and pharmaceutically acceptable polyoxyethylene-polyoxypropylene copolymers. Suitable agents which maintain or increase the conformational stability of LIF are also known to the person skilled in the art and include sugars and polyhydric alcohols.

Suitable buffers for attaining the desired pH of the composition will be known to those skilled in the art and include phosphate, citrate and acetate buffers. Preferred buffers are citrate and acetate.

Yet another aspect of the present invention contemplates a method of preparing a composition comprising Leukaemia Inhibitory Factor or a derivative or homologue thereof and which exhibits reduced deamidation and/or agglutination of LIF or a derivative or homologue over time said method comprising admixing LIF or its derivative or homologue with a stabilizing agent.

The compositions of the present invention may be suitable for administration in a variety of forms such as, but not limited to, parenteral (e.g. intravenous, intraperitoneal, intramuscular, intradermal), subcutaneous, nasal, rectal, vaginal, topical, buccal and sublingual.

The carrier must be pharmaceutically "acceptable" in the sense of being compatible with the other ingredients of the composition and not injurious to the subject. The compositions may conveniently be presented in unit dosage form and may be prepared by any methods well known in the art of pharmacy. Such methods include the step of bringing into association the active ingredient with the carrier which constitutes one or more accessory ingredients. In general, the compositions are prepared by uniformly and intimately bringing into association the active ingredient with liquid carriers or finely divided solid carriers or both, and then if necessary shaping the product.

Compositions of the present invention suitable for oral administration may be presented as a solution an aqueous or non-aqueous liquid; or as an oil-in-water liquid emulsion or a water-in-oil liquid emulsion. The active ingredient may also be presented as a bolus, electuary or paste.

Compositions suitable for topical administration in the mouth include lozenges comprising the active ingredient in a flavoured base, usually sucrose and acacia or tragacanth gum; pastilles comprising the active ingredient in an inert basis such as gelatin and glycerin, or sucrose and acacia gum; and mouthwashes comprising the active ingredient in a suitable liquid carrier.

Compositions for rectal administration may be presented as a suppository with a suitable base comprising, for example, cocoa butter.

Compositions suitable for vaginal administration may be presented as pessaries, tampons, creams, gels, pastes, foams or spray formulations containing in addition to the active ingredient such carriers as are known in the art to be appropriate.

Compositions suitable for parenteral administration include aqueous and non-aqueous isotonic sterile injection solutions which may contain anti-oxidants, buffers, bactericides and solutes which render the composition isotonic with the blood of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents. The compositions may be presented in unit-dose or multi-dose sealed containers, for example, ampoules and vials, and may be stored in a freeze-dried (lyophilised) condition

requiring only the addition of the sterile liquid carrier, for example water for injections, immediately prior to use. Extemporaneous injection solutions and suspensions may be prepared from sterile powders, granules and tablets of the kind previously described.

Preferred unit dosage compositions are those containing a daily dose or unit, daily sub-dose, as herein above described, or an appropriate fraction thereof, of the active ingredient.

It is also understood that the compositions of the present invention may also comprise one or more active agents or ingredients such a cytokines e.g. interleukins, CD antigens, colony stimulating factors, interferons and tissue necrosis factor.

It should be understood that in addition to the active ingredients particularly mentioned above, the compositions of this invention may include other agents conventional in the art having regard to the type of composition in question, for example, those suitable for oral administration may include such further agents as binders, sweeteners, thickeners, flavouring agents disintegrating agents, coating agents, preservatives, lubricants and/or time delay agents. Suitable sweeteners include sucrose, lactose, glucose, aspartame or saccharine. Suitable disintegrating agents include corn starch, methylcellulose, polyvinylpyrrolidone, xanthan gum, bentonite, alginic acid or agar. Suitable flavouring agents include peppermint oil, oil of wintergreen, cherry, orange or raspberry flavouring. Suitable coating agents include polymers or copolymers of acrylic acid and/or methacrylic acid and/or their esters, waxes, fatty alcohols, zein, shellac or gluten. Suitable preservatives include sodium benzoate, vitamin E, alpha-tocopherol, ascorbic acid, methyl paraben, propyl paraben or sodium bisulphite. Suitable lubricants include magnesium stearate, stearic acid, sodium oleate, sodium chloride or talc. Suitable time delay agents include glyceryl monostearate or glyceryl distearate.

A number of formulations of LIF were investigated in order to establish optimum conditions under which chemical and physical degradation is reduced compared to the currently employed formulation of 3.67 mg/ml in 2 mM phosphate buffer, pH 6.4-6.8.

Ion Exchange (IE), Reversed Phase (RP) and Size Exclusion (SEC) chromatography were used

to detect changes in chemical and physical degradation.

Freeze/thaw studies revealed high solubility of LIF, i.e. no aggregation, in formulations in the pH range of 4.0 - 6.0 examined, the highest being in the pH range of 4.5 to 5.5, with optimized stability at pH 5.0.

Studies of the various solutions over varying periods of storage time (0 to 8 weeks) and at a range of storage temperatures (-80 to 25°C) revealed optimum stability of the solution was achieved in a preferred pH range of 4.5 to 5.5.

The inventors examined a number of pH levels and stabilizing agents. Samples at pH 4.0, 4.5, 5.0, 5.5 and 6.0 were prepared in Examples 1 and 2, as described hereinafter, and additional stabilizing agents, Sorbitol, an isotonicity agent, and Polysorbate 80, as a non-ionic surfactant to reduce non-specific adsorption onto surfaces, including glass, were also included. NaCl was also examined as an isotonicity agent.

LIF is present in the compositions of the invention in effective amounts. Effective amounts include from 0.1 mg/ml to 100 mg/ml. Preferred effective amounts are from 10 mg/ml to 10 mg/ml. Particularly preferred amounts range from 400 mg/ml to 1000 mg/ml.

Suitable amounts of surfactant and isotonic agents may range from 0.001 to 30%. Preferably from 0.01 to 10%, even more preferably from 0.01 to 5.0%.

Particularly preferred compositions are those comprising LIF, sorbitol, polysorbate and a citrate or acetate buffer in the preferred ranges described above.

The present invention further provides for the use of a stabilizing agent in the manufacture of a composition exhibiting improved chemical and/or physical stability of Leukaemia Inhibitory Factor (LIF) or a derivative or homologue thereof.

The invention will now be described with reference to the following non-limiting Examples.

Example 1.

I. Preliminary Formulation Screening

On the basis of preliminary stability data, it was anticipated that deamidation of LIF would represent the principal pathway for degradation of solutions at neutral to slightly alkaline pH. Solution pH was, therefore, considered to be important and was a primary variable evaluated in these stability studies. Screening studies evaluating LIF stability during freeze/thaw cycling, following filtration, upon contact with vials and syringes and following temperature controlled storage were conducted in the pH range of 4 to 6 using acetate and citrate buffers at low concentrations (10 mM for each). Osmolality was controlled by the addition of sorbitol at a concentration of 5% w/v. To minimise the potential for LIF adsorption to vials, filters, and syringes, 0.01% w/v Polysorbate 80 was added to all preliminary formulations evaluated in this series of studies.

II. Analytical Methods

Three analytical methods were used to assess LIF stability upon storage. A reversed phase assay, using a standard wide pore C8 reversed phase column, was utilised for the purpose of total LIF concentration determination. The reversed phase assay was not stability indicating and therefore was not suitable for the determination of degradation products. A cation ion exchange assay was used to assess degradation products resulting from a change in the charge characteristics of the parent compound as deamidation had previously been determined to be the principal pathway for LIF degradation. A size exclusion assay was also used to detect size related changes (either cleavage, crosslinking, or aggregation) upon storage.

A. Reversed Phase (RP) Assay

Reversed phase chromatography was conducted using a wide pore C8 reversed phase column, and a trifluoroacetic acid /acetonitrile mobile phase with gradient elution. Detection was conducted at 210 nm.

B. Ion-Exchange (IEC) Assay

Ion exchange chromatography was conducted using a cation exchange column, pH 7 phosphate buffer and a salt gradient. Detection was conducted at 280 nm.

C. Size Exclusion (SEC) Assay

Size exclusion chromatography was conducted using a dextrose based size exclusion column with a molecular weight range of 10 to 300 Daltons. The mobile phase was a pH 7.2 phosphate buffer and detection was conducted at 210 nm.

III. Method Validation

A. Reversed Phase (RP) Assay

Using the defined RP conditions, LIF eluted as a sharp, symmetrical peak with a retention time of approximately 37 min as shown in Figure 1. The RP assay was used for quantitation of total LIF only as the method was not selective for LIF in the presence of degradation (deamidation or dimeric) products.

Calibration curves for total peak area versus LIF concentration were prepared with each set of analyses in the concentration range of 0.2 and 1.0 mg/ml LIF.

Precision was determined from the coefficient of variation (CV, %) for the total peak area obtained for replicate injections of standard solutions prepared at 0.4 and 1.0 mg/ml. Accuracy was determined by comparison of the total peak area for these standard solutions to a separately prepared calibration curve and was expressed as the percentage deviation from the nominal concentration. Results for accuracy and precision with the RP assay are shown in Table 1. A summary of the RP calibration curves is shown in Table 2.

B. Ion-Exchange (IEC) Assay

Using the defined IEC conditions, LIF eluted as a slightly tailing peak with a retention time of approximately 13 min as shown in Figure 2. Separation of the main LIF peak from degradation (deamidation) products formed following storage was observed during the course of the studies. The actual identity of the degradation products (i.e. site of deamidation) was not

determined in these studies.

Calibration curves for total peak area (main peak plus degradation products) versus LIF concentration were prepared with each set of analyses in the concentration range of 0.2 and 1.0 mg/ml LIF. Calibration curves were linear in this range when 100 µl was injected onto the column.

Precision was determined from the coefficient of variation (CV, %) for the total peak area obtained for replicate injections of standard solutions prepared at 0.4 and 1.0 mg/ml. Accuracy was determined by comparison of the total peak area for these standard solutions to a separately prepared calibration curve and was expressed as the percentage deviation from the nominal concentration. Results for precision and accuracy for the IEC assay are shown in Table 3. A summary of the IEC calibration curves over the course of the study is shown in Table 4.

C. Size Exclusion (SEC) Assay

Using the defined SEC conditions, LIF eluted as a sharp, symmetrical peak with a retention time of approximately 26 min as shown in Figure 3. The method separated monomeric LIF from dimeric LIF which eluted at approximately 21 min, but was not selective for other degradation (deamidation) products which eluted as monomeric LIF.

Calibration curves for total peak area (main peak plus degradation products) versus LIF concentration were prepared with each set of analyses in the concentration range of 0.2 and 1.0 mg/ml LIF.

Precision was determined from the coefficient of variation (CV, %) for the total peak area obtained for replicate injections of standard solutions prepared at 0.4 and 1.0 mg/ml. Accuracy was determined by comparison of the total peak area for these standard solutions to a separately prepared calibration curve and was expressed as the percentage deviation from the nominal concentration. Results for precision and accuracy for the SEC assay are shown in Table 5. A summary of the SEC calibration curves is shown in Table 6.

IV. Buffer Composition

All LIF samples were prepared by dilution of stock LIF solution containing 3.67 mg/ml LIF in 2 mM phosphate buffer, pH 6.42 to give the desired final LIF concentration (either 0.4 or 1.0 mg/ml) and composition of buffer components. In these studies, the final composition of each solution contained 10 mM buffer (either acetate or citrate), 5% w/v sorbitol and 0.01% w/v Polysorbate 80. Samples differed in the final concentration of phosphate buffer (present from the original stock LIF solution) depending on the dilution factor. The 0.4 mg/ml LIF solutions contained 0.22 mM residual phosphate while the 1.0 mg/ml LIF solutions contained 0.54 mM residual phosphate. The composition of each buffer was as follows:

A. Acetate Buffer for 0.4 mg/ml LIF Formulations

Solution A: 11.22 mM sodium acetate trihydrate (Merck #1.06267)
5.61% w/v sorbitol (Sigma Chemicals #S1876)
0.0112% w/v Polysorbate 80 (Sigma Chemicals #P1754)

Solution B: 11.22 mM glacial acetic acid (Sigma Chemicals #A6283)
5.61% w/v sorbitol (Sigma Chemicals #S1876)
0.0112% w/v Polysorbate 80 (Sigma Chemicals #P1754)

Solutions A and B were mixed to give a final pH of 4.0 or 4.5. Formulations were prepared by combining 0.109 parts stock LIF solution and 0.891 parts buffer to give a final LIF concentration of 0.4 mg/ml, a final buffer concentration of 10 mM, a final sorbitol concentration of 5% w/v and a final Polysorbate 80 concentration of 0.01% w/v.

B. Acetate Buffer for 1.0 mg/ml LIF Formulations

Solution A: 13.75 mM sodium acetate trihydrate (Merck #1.06267)
6.88% w/v sorbitol (Sigma Chemicals #S1876)
0.0138% w/v Polysorbate 80 (Sigma Chemicals #P1754)

Solution B: 13.75 mM glacial acetic acid (Sigma Chemicals #A6283)
6.88% w/v sorbitol (Sigma Chemicals #S1876)

0.0138% w/v Polysorbate 80 (Sigma Chemicals #P1754)

Solutions A and B were mixed to give a final pH of 4.0 or 4.5. Formulations were prepared by combining 0.272 parts stock LIF solution and 0.728 parts buffer to give a final LIF concentration of 1.0 mg/ml, a final buffer concentration of 10 mM, a final sorbitol concentration of 5% w/v and a final Polysorbate 80 concentration of 0.01% w/v.

C. Citrate Buffer for 0.4 mg/ml LIF Formulations

Solution A: 11.22 mM sodium citrate dihydrate (Merck #1.06448)
5.61% w/v sorbitol (Sigma Chemicals #S1876)
0.0112% Polysorbate 80 (Sigma Chemicals #P1754)

Solution B: 11.22 mM citric acid monohydrate (Merck #1.00244)
5.61% w/v sorbitol (Sigma Chemicals #S1876)
0.0112% Polysorbate 80 (Sigma Chemicals #P1754)

Solutions A and B were mixed to give a final pH of 5.0, 5.5, or 6.0. Formulations were prepared by combining 0.109 parts stock LIF solution and 0.891 parts buffer to give a final LIF concentration of 0.4 mg/ml, a final buffer concentration of 10 mM, a final sorbitol concentration of 5% w/v and a final Polysorbate 80 concentration of 0.01% w/v.

D. Citrate Buffer for 1.0 mg/ml LIF Formulations

Solution A: 13.75 mM sodium citrate (Merck #1.06448)
6.88% w/v sorbitol (Sigma Chemicals #S1876)
0.0138% w/v Polysorbate 80 (Sigma Chemicals #P1754)

Solution B: 13.75 mM citric acid (Merck #1.00244)
6.88% w/v sorbitol (Sigma Chemicals S1876)
0.0138% w/v Polysorbate 80 (Sigma Chemicals P1754)

Solutions A and B were mixed to give a final pH of 5.0, 5.5, or 6.0. Formulations were

prepared by combining 0.272 parts stock LIF solution and 0.728 parts buffer to give a final LIF concentration of 1.0 mg/ml, a final buffer concentration of 10 mM, a final sorbitol concentration of 5% w/v and a final Polysorbate 80 concentration of 0.01% w/v.

Table 7 displays pH and osmolality (obtained using a Fiske One-Ten Osmometer) values for 0.4 and 1.0 mg/ml LIF samples prepared using the above buffer systems.

V. Freeze/Thaw Cycling

A. Sample Preparation and Methods

LIF samples were prepared by dilution of stock LIF (3.67 mg/ml in 2 mM phosphate buffer, pH 6.8) with acetate or citrate buffer containing sorbitol and polysorbate 80 to give a final buffer concentration of 10 mM, a theoretical pH of 4.0, 4.5, 5.0, 5.5, or 6.0, a final sorbitol concentration of 5% w/v, a final polysorbate 80 concentration of 0.01% w/v and a final LIF concentration of 1 mg/ml (see Section IV). The final pH of each sample was essentially the same as predicted by theory. Solutions (3 ml) were filtered through 0.22 m sterile filters (Millex GV) with the first 0.5 ml aliquot from the filter being retained as a separate sample for the preliminary determination of filter adsorption. Subsequent 0.5 ml aliquots were filtered into sterile 2 ml glass vials and capped with sterile rubber/teflon lined caps and crimped. One vial for each formulation was analysed on the day of preparation and all other vials were stored at -80°C. On each of 5 days, all vials were thawed and one vial of each formulation was centrifuged and an aliquot taken for dilution (in this study, all samples were analysed at a LIF concentration of 0.1 mg/ml) and analysis by RP, IEC, and SEC.

A 0.1 mg/ml standard solution was prepared by diluting the LIF stock solution with 2 mM phosphate buffer, pH 6.42 containing 0.01% polysorbate 80. This standard solution was stored at 4°C for a total of 6 days and analysed along with each sample set.

B. Results

Figure 4 represents the individual peak areas for samples at each pH with concentration being expressed as a percentage of the initial concentration measured by each of the three methods.

While there was some variability in the individual results (most likely due to the dilution step prior to analysis), there were no trends which would indicate loss of LIF upon freeze/thaw cycling.

Figure 5 illustrates the average concentration (as a percentage of the initial concentration) over 5 freeze/thaw cycles for each of the different pH values.

VI. Long Term Stability at -80°C, -20°C, 8°C and 25°C

A. Preparation of Samples for Storage at -80°C and -20°C

Five LIF formulations were prepared by dilution of stock LIF (3.67 mg/ml in 2 mM phosphate buffer, pH 6.42) with acetate or citrate buffer containing sorbitol and polysorbate 80 to give a final LIF concentration of 0.4 mg/ml or 1.0 mg/ml, a final buffer concentration of 10 mM, a final sorbitol concentration of 5% w/v and a final polysorbate 80 concentration of 0.01% w/v (see Section V). The theoretical pH values were pH 4.0 (acetate buffer), 4.5 (acetate buffer), and 5.0 (citrate buffer). The final pH of each sample was essentially the same as predicted by theory.

Under aseptic conditions in a laminar flow cabinet, the formulations were sterile filtered using 0.22 m Millex GV (Millipore) filters. The first 1.0 ml of each filtrate was set aside and the vial marked accordingly (previous studies identified that approximately 1 ml was required to saturate the filter binding sites using Millex GV filter units). The remaining volume was filtered into a sterile 50 ml polypropylene tube. Aliquots of each formulation (1.15 ml/vial) were transferred using a multiple dispensing Eppendorf pipette with sterile tips into heat sterilised 2 ml glass vials and capped with sterile teflon lined rubber caps which were then crimped. Vials were labelled and duplicate vials of each formulation were retained for the initial analysis. The remaining vials were stored at either -80°C or -20°C.

B. Preparation of Samples for Storage at 8°C and 25°C

Five LIF formulations were prepared by a dilution of stock LIF (3.67 mg/ml in 2 mM phosphate buffer, pH 6.42) with acetate or citrate buffer containing sorbitol and polysorbate 80 to give a final LIF concentration of 0.4 mg/ml or 1.0 mg/ml, a final buffer concentration of 10 mM, a

final sorbitol concentration of 5% w/v and a final polysorbate 80 concentration of 0.01% w/v. The theoretical pH values were pH 4.0 (acetate buffer), 4.5 (acetate buffer), and 5.0 (citrate buffer). The final pH of each sample was essentially the same as predicted by theory.

Formulations were filtered and filled into vials as described for the -80°C and -20°C samples. Samples were stored in temperature controlled incubators at either 8°C or 25°C. Incubators were checked daily to ensure the correct temperature was maintained.

C. Sample Analysis

All LIF samples were analysed undiluted according to the methods described in Section III. LIF standards at concentrations of 0.2, 0.4, 0.7 and 1.0 mg/ml were prepared from stock LIF (3.67 mg/ml in 2 mM phosphate buffer) by diluting with 2 mM phosphate buffer, pH 6.42 containing 0.01% w/v polysorbate 80. These standards were prepared fresh at the beginning of each set of analyses and were analysed along with the samples at the start and end of each analytical run.

At each time point, 2 vials were withdrawn from the freezers or incubators and approximately 200 µl was removed from each using a sterile 1 ml syringe and a sterile needle. These aliquots were placed into polypropylene autosampler vials and sealed with caps containing self-sealing septa to allow repeat injections from the same vial without evaporation.

Autosampler vials were transferred to the autosampler where they were maintained at 4°C throughout the three analytical runs. The same sample and standard autosampler vials were used for each of the three analyses with the RP (10 µl injection volume) being conducted first, followed by the IEC (100 µl injection volume) and then the SEC (10 µl injection volume). The complete RP run took approximately 32 hours, and the IEC and SEC runs took approximately 25 hours each. It was assumed that any further degradation over this storage time in the autosampler would be minimal (standard solutions at pH 6.42 stored under the same conditions showed no change over the complete analytical period). Samples were analysed in the following order:

Blank x 2

Standards 0.2 mg/ml, 0.4 mg/ml, 0.7 mg/ml and 1.0 mg/ml

Blank		
0.4 mg/ml	Acetate pH 4.0	x2
	Acetate pH 4.5	x2
	Citrate pH 5.0 x2	
1.0 mg/ml	Acetate pH 4.5	x2
	Citrate pH 5.0 x2	
Blank		
Standards	0.2 mg/ml, 0.4 mg/ml, 0.7 mg/ml and 1.0 mg/ml	

Selected samples were also analysed for particulates using a Malvern Instruments Zetasizer 3000 particle size instrument. Samples were withdrawn from the storage vials using a syringe and placed in the sample cuvette. Samples were counted for 120 sec using a 200 m pinhole (to obtain the maximum signal), 90° scattering angle, and scattering source at 633 nm using a 10 mW He-Ne ion laser.

D. Results

Data pertaining to solution pH, LIF concentration in mg/ml (determined by comparison to LIF standard solutions), and the area % of the main peak relative to the total peak area for all LIF related peaks in the chromatogram analysed using the three chromatographic methods are shown in Tables 8 through 17. None of the samples showed significant shifts in pH over the storage period.

1. Ion Exchange

Figures 6 through 15 illustrate IEC chromatograms for samples stored in each of the different buffer systems at 8 and 25°C. Two main products were evident for samples prepared in pH 4.0 and 4.5 buffers (eluting at approximately 9 and 10 min) whereas a single main product (eluting at approximately 10 min) was seen in the pH 5.0 samples. At each pH, there was evidence of several minor degradation products in the ion exchange chromatograms, however, due to inadequate resolution between the different products, the exact number of products could not be determined. Representative chromatograms for samples stored at -80 and -20°C are not

shown as they were similar to the chromatograms at the higher temperatures with degradation products being present at significantly reduced levels.

The IEC results for samples stored at -80, -20, 8 and 25°C are shown graphically in Figures 16 through 18 with the main LIF peak plotted as a percentage of the total area for all LIF related peaks in the chromatogram as a function of storage time. The data illustrate the dependence of LIF stability on pH and temperature. The relative stability under each storage condition was similar for the 0.4 and 1.0 mg/ml formulations. The pH 4.0 samples displayed significant variability between the different time points at 8 and 25°C. Re-analysis of selected samples gave similar results to the original values. There was also evidence of degradation at pH 4.0 and 4.5 following storage at -20°C and -80°C. The stability was greatly improved at pH 5 in comparison to pH 4 and 4.5. After 55 days storage at 8°C, approximately 97% of the total peak area was present as the main LIF peak. Following storage at 25°C for 55 days, this value was reduced to approximately 78%. Samples prepared at pH 5 and stored at -80 or -20°C for up to 84 days showed no significant evidence of degradation.

2. Reversed Phase

Representative RP chromatograms are not included as all displayed essentially the same elution characteristics (see Figure 1). In all cases, the chromatograms showed the presence of only one main peak eluting at approximately 36 min.

The RP results for samples stored at -80, -20, 8 and 25°C, wherein the measured concentration was plotted as a function of storage time, illustrated the absence of significant change in the measured concentration over the storage period for each of the buffer and storage conditions utilised.

3. Size Exclusion

SEC chromatograms for the samples as all displayed essentially the same elution characteristics (see Figure 3). In all cases, the chromatograms showed the presence of one main peak eluting

at approximately 26 min and a minor peak eluting at approximately 21 min.

The SEC results for samples stored at -80, -20, 8 and 25°C wherein the measured concentration was plotted as a function of storage time, illustrated the absence of significant change in the measured concentration over the storage period for each of the buffer and storage conditions utilised. Using the SEC method, there was no evidence of chain cleavage or crosslinking under the storage conditions studied.

4. Particle Size Analysis

Samples stored for 56 days at -80 and -20°C and for 41 days at 8 and 25°C were analysed for particulates using a laser light scattering instrument. All of the samples analysed displayed a count rate of "0 kCps" which effectively means that the samples contained no particulates (i.e. no signal was measurable).

VII. Summary

These studies demonstrated no notable loss of LIF following freeze thaw cycling of 1.0 mg/ml LIF solution formulations prepared in acetate or citrate buffers (pH 4 to 6) containing 5% w/v sorbitol and 0.01% w/v polysorbate 80. There was no significant loss of LIF on 0.2 m Sartorius Minisart filters when formulations were prepared at either 0.4 or 1.0 mg/ml in pH 5.0 or 5.5 citrate buffers containing 5% w/v sorbitol and 0.01% w/v polysorbate 80. For the pH 5.0 and 5.5 formulations, there was also no evidence of loss of LIF on the proposed vials, stoppers, or syringes.

At -80°C, there was no significant change in LIF concentrations measured by RP, IEC and SEC methods following storage for 84 days in the pH range of 4 to 5. At -20°C over the same time period, there was evidence of degradation for formulations prepared at pH 4 and analysed by IEC, but the remaining formulations were stable under these storage conditions. Generally, 0.4 and 1.0 mg/ml LIF formulations displayed similar stability characteristics under each of the conditions investigated. Formulations prepared at pH 5 were found to be stable for up to 8 weeks when stored at 8°C with minimal loss of the parent compound (~1%) shown by IEC and

no loss shown by RP or SEC.

Table 1. Precision and Accuracy for the RP Assay

Nominal Conc. (mg/ml)	Total Peak Area	Measured Conc. (mg/ml)	Precision (CV, %)	Accuracy (% deviation)
0.4	14.213	0.391	0.44 (n=5)	-2.16
0.4	14.356	0.395		-1.21
0.4	14.361	0.395		-1.17
0.4	14.322	0.394		-1.43
0.4	14.255	0.392		-1.88
1.0	38.002	1.029	0.39 (n=5)	2.92
1.0	38.170	1.034		3.37
1.0	38.327	1.038		3.79
1.0	38.344	1.038		3.84
1.0	38.077	1.031		3.12

Table 2. Summary of RP Calibration Curves Over the Course of the Study

	Slope	Intercept
	33.460	-1.755
	32.900	-0.312
	34.491	-1.040
	32.648	-0.137
	32.865	1.006
	32.865	0.556
	33.705	1.092
	34.617	0.535
	35.920	0.113
	35.666	-0.014
	37.294	-0.382
mean	34.221	-0.030
SD	1.529	
CV,%	4.469	

Table 3 Precision and Accuracy for the IEC Assay

Nominal Conc (mg/ml)	Total Peak Area	Measured Conc. (mg/ml)	Precision (CV, %)	Accuracy (% deviation)
0.4	8.310	0.397	0.68 (n=5)	-0.86
0.4	8.260	0.398		-0.62
0.4	8.265	0.399		-0.30
0.4	8.232	0.396		-1.10
0.4	8.234	0.403		0.65
1.0	21.929	1.007	0.41 (n=5)	0.70
1 0	21.910	1.005		0.51
1.0	21.918	1.008		0.77
1.0	21.901	1.004		0.35
1.0	21.870	1.014		1.43

Table 4. Summary of IEC Calibration Curves Over the Course of the Study

	Slope	Intercept
	2.953	-0.002
	3.111	-0.038
	3.104	-0.048
	2.983	-0.019
	2.987	-0.020
	3.005	-0.018
	2.942	-0.012
	3.064	-0.055
	3.005	-0.018
	3.034	-0.036
	3.137	-0.099
mean	3.030	-0.033
SD	0.066	—
CV,%	2.180	—

Table 5. Precision and Accuracy for the SEC Assay

Nominal Conc (mg/ml)	Total Peak Area	Measured Conc. (mg/ml)	Precision (CV, %)	Accuracy (% deviation)
0.4	8.310	0.396	0.39 (n=5)	-0.98
0.4	8.260	0.394		-1.48
0.4	8.265	0.394		-1.46
0.4	8.232	0.393		-1.84
0.4	8 234	0.393		-1.86
1.0	21.929	1.002	0.11 (n=5)	0.23
1.0	21.910	1.001		0.11
1.0	21.918	1.001		0.15
1.0	21.901	1.000		0.07
1.0	21.870	0.999		-0.05

Table 6. Summary of SEC Calibration Curves Over the Course of the Study

	Slope	Intercept
	21.332	0.202
	21.278	0.166
	22.351	0.230
	21.672	0.054
	20.810	0.419
	21.561	0 130
	21.845	0.074
	21 883	-0.090
	21.963	0.158
	21.794	-0.003
	22.558	-0.474
mean	21.732	0.079
SD	0.491	—
CV,%	2.258	—

Table 7. pH and Osmolality of AM424 Formulations

buffer / theoretical pH	AM424 conc. (mg/ml)	measured pH	osmolality (mOsm/kg)
Acetate / pH 4.0	0.4	3.95	297
Acetate / pH 4.5	0.4	4.48	297
Citrate / pH 5.0	0.4	4.94	303
Acetate / pH 4.5	1.0	4.47	294
Citrate / pH 5.0	1.0	4.96	305

Table 8 Summary of 0.4 mg/ml, pH 4.0 AM424 Formulation Stability Following Storage at 8°C and 25°C

measured pH	buffer	Nominal AM424 Conc. (mg/ml)	Storage Temp (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
4 03	acetate	0 4	8	0	0.40, 0.39	100, 100	0.37, 0.37	98.9, 99.0	0.39, 0.40	98 8, 98 7
----				7	0.40, 0.40	100, 100	0.35, 0 34	91.7, 87.2	0.39, 0.40	97 6, 97 5
4 07				13	0.39, 0.39	100, 100	0.33, 0.37	90.8, 92.8	0.40, 0.40	98 9, 98 9
----				19	0.40, 0.40	100, 100	0 34, 0 33	89.7, 86 8	0.38, 0.38	98 8, 99 0
4 06				27	0.40, 0.40	100, 100	0 33, 0 33	84.6, 83 7	0.40, 0.40	98.9, 98 9
4 06				41	0.40, 0.40	100, 100	0 34, 0 35	86.9, 88 2	0.40, 0.41	98.9, 98 9
4 16				55	0.40, 0.41	100, 100	0 34, 0.33	89.2, 83.0	0.40, 0.40	99 0, 99 0
4 03	acetate	0.4	25	0	0.40, 0.39	100, 100	0.37, 0 37	98.9, 99.0	0.39, 0.40	98 8, 98 7
----				7	0.39, 0.39	100, 100	0.33, 0.36	85.1, 91.5	0.39, 0.40	97.3, 97 4
4 06				13	0.40, 0.39	100, 100	0.28, 0.30	74.7, 80.7	0.39, 0.41	99.2, 99.1
----				19	0.40, 0.39	100, 100	0.31, 0 32	78.3, 80.3	0.38, 0.38	99.0, 99 2
4 07				27	0.40, 0.40	100, 100	0.29, 0 30	73.3, 74.5	0.40, 0.40	99.4, 99 2
4 09				41	0.40, 0.40	100, 100	0.31, 0 31	76.1, 77.8	0.41, 0.41	99.2, 99 2
4.12				55	0.41, 0.40	100, 100	0.25, 0.24	62.6, 59.8	0.40, 0.40	99.3, 99.7

Underlined values represent repeat analyses

Table 9 Summary of 0.4 mg/ml, pH 4.0 AM424 Formulation Stability Following Storage at -80°C and -20°C

measured pH	buffer	Nominal AM424 Conc (mg/ml)	Storage Temp (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
3 95	acetate	0.4	-80	0	0.41, 0.40	100, 100	0 38, 0.38	98.4, 98.6	0.40, 0.40	98.9, 98.5
3 98				28	0.41, 0.41	100, 100	0.38, 0.39	97.8, 98.8	0.39, 0.40	98.2, 98 0
3 99				56	0.41, 0.41	100, 100	0.37, 0.38	96.6, 98.9	0.39, 0.39	98.3, 98 3
4.05				84	0.43, 0.42	100, 100	0.40, 0.38	98.6, 99.1	0.41, 0.41	99.3, 98.6
3.95	acetate	0.4	-20	0	0.41, 0.40	100, 100	0.38, 0.38	98.4, 98.6	0.40, 0.40	98.9, 98.5
3.95				28	0.41, 0.42	100, 100	0.38, 0 39	96.9, 97.8	0.40, 0.40	98.7, 98 5
4 04				56	0.40, 0.41	100, 100	0.36, 0.36	94.2, 93.7	0.40, 0.40	99.0, 98 9
4.03				84	0.42, 0.43	100, 100	0.38, 0.38	92.5, 93.1	0.42, 0.41	99.2, 98 9

Table 10. Summary of 0.4 mg/ml, pH 4.5 AM424 Formulation Stability Following Storage at 8°C and 25°C

measured pH	buffer	Nominal AM424 Conc. (mg/ml)	Storage Temp. (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
4 52	acetate	0.4	8	0	0.39, 0.39	100, 100	0.36, 0.36	99.0, 98.9	0.39, 0.38	98.8, 98 8
----				7	0.38, 0.38	100, 100	0.37, 0.36	95.4, 95.6	0.38, 0.39	97.7, 97.8
4 53				13	0.38, 0.38	100, 100	0 38, 0.36	98.3, 95.6	0.39, 0.38	99.0, 99.0
----				19	0.38, 0.38	100, 100	0 37, 0.35	97.8, 93.3	0.38, 0.38	98.2, 98.2
4 53				27	0.38, 0.38	100, 100	0 35, 0 36	90.8, 94.1	0.39, 0.39	98.9, 98 8
4 51				41	0.39, 0.39	100, 100	0 37, 0.36	95.3, 94.2	0.39, 0.39	98.9, 98 8
4 59				55	0.40, ----	100, ----	0 35, 0.33	89.6, 85.9	0.39, 0.39	99.0, 98 9
4 52	acetate	0 4	25	0	0.39, 0.39	100, 100	0.36 0.36	99.0, 98.9	0.39, 0.39	98.8, 98 8
----				7	0.38, 0.38	100, 100	0.36, 0.34	94.7, 88.8	0.39, 0.39	98.1, 98.2
4.52				13	0.39, 0.38	100, 100	0.33, 0.35	86.8, 91.0	0.39, 0.38	99.0, 99 0
----				19	0.38, 0.38	100, 100	0.31, 0.30	82.0, 80.0	0.38, 0.38	99.1, 99.0
4.52				27	0.38, 0.38	100, 100	0.30, 0.29	75.8, 73.5	0.39, 0.39	99.1, 99 2
4.53				41	0.40, 0.40	100, 100	0 28, 0.28	71.2, 71.1	0.39, 0.39	99.2, 99 3
4.55				55	0.39, 0.40	100, 100	0.22, 0.24	53.4, 59.1	0.39, 0.39	99.3, 99.4

Underlined values represent repeat analyses

Table 11. Summary of 0.4 mg/ml, pH 1.5 AM424 Formulation Stability Following Storage at -80°C and -20°C

measured pH	buffer	Nominal AM424 Conc. (mg/ml)	Storage Temp. (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
4.48	acetate	0.4	-80	0	0.40, 0.40	100, 100	0.39, 0.40	99.0, 98.9	0.40, 0.40	98.9, 98.8
4.49				28	0.41, 0.40	100, 100	0.39, 0.38	98.7, 98.7	0.40, 0.39	98.1, 98.0
4.49				56	0.40, 0.40	100, 100	0.38, 0.38	98.5, 98.6	0.39, 0.39	98.4, 98.2
4.55				84	0.42, 0.42	100, 100	0.40, 0.40	98.6, 98.4	0.42, 0.42	98.5, 98.5
4.48	acetate	0.4	-20	0	0.40, 0.40	100, 100	0.39, 0.40	99.0, 98.9	0.40, 0.40	98.9, 98 8
4.47				28	0.41, 0.41	100, 100	0 39, 0.38	98.8, 96.9	0.40, 0.40	98.4, 98 6
4.52				56	0.40, 0.40	100, 100	0.39, 0.38	98.5, 97.3	0.40, 0.39	98.6, 98 7
4.53				84	0.42, 0.42	100, 100	0.40, 0 40	96.4, 96.5	0.42, 0.42	99.0, 99 0

Table 12 Summary of 1.0 mg/ml, pH 4.5 AM424 Formulation Stability Following Storage at 8°C and 25°C

measured pH	buffer	Nominal AM424 Conc. (mg/ml)	Storage Temp. (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
4 54	acetate	1.0	8	0	0.99, 0.99	100, 100	0.96, 0.96	98.5, 98.6	0.99, 0.99	98.6, 98 2
----				7	0.98, 0.98	100, 100	0.98, 0.99	96.9, 979	0.99, 0.99	98.6, 98 6
4 57				13	0.98, 0.99	100, 100	0.94, 0.96	96.2, 97.6	0.98, 0.98	98.8, 98.7
----				19	0.98, 1.00	100, 100	0.96, 0.94	97.5, 95.6	1.00, 0.99	98.6, 98 8
4 56				27	0.99, 1.00	100, 100	0.94, 0.89	97.0, 90.1	1.00, 1.00	98.6, 98.8
4 55				41	0.98, 0.99	100, 100	0 88, 0.90	90.3, 92.1	0.98, 0.98	98.9, 98 9
4 61				55	0.99, 1.00	100, 100	0 90, 0.85	91.2, 86.1	0.99, 0.99	98.9, 98 9
4 54	acetate	1 0	25	0	0.99, 0.99	100, 100	0.96, 0.96	98.5, 98.6	0.99, 0.99	98.6, 98 2
----				7	0.99,0.99	100, 100	0.92, 0.94	91.4, 92.7	0.99, 0.99	98.9, 98 9
4.57				13	1.00, 0.99	100, 100	0.82, 0 86	83.6, 88.6	0.98, 0.98	99.0, 99 0
----				19	1.00, 1.00	100, 100	0.84, 0.81	83.7, 80.9	1.00, 1.00	98.9, 98 9
4 57				27	1.00, 1.00	100, 100	0.78, 0.81	77.1, 79.3	1.00, 1.00	99.0, 99 0
4.59				41	0.99, 0.99	100, 100	0.68, 0.65	66.4, 63.9	0.98, 0.98	98.9, 99 1
4.61				55	1.00, 0.99	100, 100	0.59, 0.60	56.7, 58.9	0.99, 0.99	99.1, 99.1

Underlined values represent repeat analyses

Table 13. Summary of 1.0 mg/ml, pH 4.5 AM424 Formulation Stability Following Storage at -80°C and -20°C

measured pH	buffer	Nominal AM424 Conc. (mg/ml)	Storage Temp. (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
4.47	acetate	1.0	-80	0	1.00, 1.00	100, 100	0.97, 0.98	98.9, 98.7	0.99, 0.99	98.7, 98.6
4.47				28	1.00, 1.00	100, 100	0.97, 0.97	98.4, 98.3	0.99, 0.99	98.4, 98.4
4.50				56	0.99, 0.98	100, 100	0.96, 0.95	98.5, 98.3	0.97, 0.97	98.5, 98.6
4.53				84	1.00, 1.00	100, 100	0.96, 0.96	98.3, 98.5	0.98, 0.98	98.6, 98.4
4.47	acetate	1.0	-20	0	1.00, 1.00	100, 100	0.97, 0.98	98.9, 98.7	0.99, 0.99	98.7, 98.6
4.48				28	1.00, 0.99	100, 100	0.98, 0.97	98.3, 97.4	1.00, 0.99	98.5, 98.6
4.50				56	0.98, 0.99	100, 100	0.94, 0.96	97.4, 98.0	0.98, 0.98	98.6, 98.5
4.51				84	0.99, 0.99	100, 100	0.95, 0.97	96.9, 98.4	0.99, 0.99	98.7, 98.5

Table 14 Summary of 0.4 mg/ml, pH 5.0 AM424 Formulation Stability Following Storage at 8°C and 25°C

measured pH	buffer	Nominal AM424 Conc (mg/ml)	Storage Temp. (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
5 02	citrate	0 4	8	0	0.38, 0.37	100, 100	0.36, 0.36	98.8, 98.7	0.38, 0.38	98.6, 98.4
----				7	0.37, 0.37	100, 100	0 38, 0 38	98.4, 98.4	0.37, 0.37	98.1, 98 1
5 03				13	0.37, 0.37	100, 100	0 38, 0 38	98.4, 98.4	0.38, 0.38	98.6, 98 7
----				19	0 37, 0.37	100, 100	0 37, 0 37	98.4, 98.5	0.37, 0.37	98.3, 98 0
5 06				27	0.38, 0.38	100, 100	0.38, 0 37	98.4, 98.5	0.38, 0.38	98 6, 98 6
5 04				41	0.39, 0.39	100, 100	0 38, 0 37	97 8, 97.9	0.38, 0.38	98.7, 98 7
5 07				55	0.39, 0.39	100, 100	0.37, 0.37	97.7, 97.5	0.39, 0.38	98.8, 98 7
5 02	citrate	0 4	25	0	0.38, 0.37	100, 100	0.36, 0.36	98.8, 98.7	0 38, 0.38	98.6, 98.4
----				7	0 37, 0.37	100, 100	0.37, 0.36	97.0, 97.0	0.38, 0.38	98.7, 98 5
5 05				13	0.38, 0.37	100, 100	0.36, 0 37	95.4, 95.1	0.37, 0.37	98.7, 98 7
----				19	0.37, 0.37	100, 100	0 35, 0.35	93.8, 93.9	0.38, 0.38	98.8, 98 8
5 05				27	0.38, 0.38	100, 100	0.34, 0.34	92.0, 91.8	0.39, 0.39	99.3, 99 0
5 06				41	0.39, 0.39	100, 100	0 33, 0 34	87 0, 87.4	0.38, 0.38	99.1, 99 1
5 03				55	0.39, 0.39	100, 100	0.30, 0.30	77.8, 77.9	0.39, 0.39	99.0, 98.9

Underlined values represent repeat analyses

Table 15. Summary of 0 4 mg/ml, pH 5.0 AM424 Formulation Stability Following Storage at -80°C and -20°C

measured pH	buffer	Nominal AM424 Conc. (mg/ml)	Storage Temp (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
4 94	citrate	0.4	-80	0	0.41, 0.40	100, 100	0 40, 0.40	98.9, 98.8	0.39, 0.39	98.8, 98 7
4 98				28	0.41, 0.41	100, 100	0 39, 0.39	98.5, 98.5	0.39, 0.39	98.3, 98.3
4 98				56	0.40, 0.40	100, 100	0 38, 0.38	98.6, 98.6	0.38, 0.38	98.4, 98 3
5 00				84	0.42, 0.42	100, 100	0.40, 0.40	98.7, 98.4	0.41, 0.41	98.5, 98 5
4 94	citrate	0.4	-20	0	0.41, 0.41	100, 100	0.40, 0.40	98.9, 98.8	0.39, 0.39	98.8, 98.7
4 95				28	0.41, 0.41	100, 100	0 39, 0 39	98.5, 98.5	0.40, 0.40	98.5, 98 6
4 96				56	0.40, 0.40	100, 100	0 38, 0 39	98.4, 98.6	0.39, 0.39	98.6, 98 6
4 97				84	0.42, 0.42	100, 100	0 41, 0.41	98.6, 98.7	0.41, 0.41	99.0, 98.8

Table 16. Summary of 1.0 mg/ml, pH 5.0 AM424 Formulation Stability Following Storage at 8°C and 25°C

measured pH	buffer	Nominal AM424 Conc. (mg/ml)	Storage Temp (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
5 00	citrate	1.0	8	0	0.98, 0.98	100, 100	0 95, 0 95	98.5, 98.5	0.97, 0.97	98.2, 98 1
----				7	0.98, 0.98	100, 100	0 99, 0 99	98.5, 98.5	0.97, 0.98	98.5, 98.5
5 05				13	0.97, 0.97	100, 100	0.94, 0 94	98.1, 98.2	0.96, 0.96	98 2, 98 0
----				19	0.99, 0.99	100, 100	0.95, 0 95	98.1, 98.0	0.98, 0.98	98.5, 98 6
5 02				27	0.98, 0.99	100, 100	0 99, 0 98	98.0, 98.1	0.98, 0.98	98.6, 98 6
5 04				41	0.96, 0.96	100, 100	0.94, 0 94	97.5, 97.6	0.95, 0.96	98 7, 98 6
5 04				55	0.98, 0.98	100, 100	0.94, 0 94	97.0, 97.2	0.97, 0.98	98.6, 98.8
5 00	citrate	1 0	25	0	0.98, 0.98	100, 100	0.95, 0 95	98.5, 98.5	0.97, 0.97	98.2, 98 1
----				7	0.97, 0.97	100, 100	0.97, 0 97	97.0, 97.0	0.98, 0.98	98.8, 98.6
5 06				13	0.98. 0.97	100, 100	0.92, 0.91	94.6, 94.7	0.97, 0.97	98.8, 98.8
----				19	0.99, 1.00	100, 100	0.90, 0 89	92.2, 92.3	0.98, 0.98	98.8, 98 6
5 05				27	0.99, 0.99	100, 100	0.91, 0.91	90.3, 90.3	0.99, 0.98	98 8, 98.8
5 06				41	0.97, 0.97	100, 100	0 80, 0.80	83.0, 83.0	0.96, 0.96	98.6, 98 7
5 00				55	0.99, 0.98	100, 100	0.76, 0.76	77.7, 78.0	0.99, 0.97	99.0, 98 7

Underlined values represent repeat analyses

Table 17. Summary of 1.0 mg/ml, pH 5 0 AM424 Formulation Stability Following Storage at -80°C and -20°C

measured pH	buffer	Nominal AM424 Conc (mg/ml)	Storage Temp (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
4 96	citrate	1.0	-80	0	1.00, 1.00	100, 100	0.98, 0 99	98.8, 98.8	0.98, 0.98	98.1, 98.1
4 97				28	1.00, 0.99	100, 100	0.96, 0.96	98.2, 98.1	0.98, 0.98	98.4, 98.4
4 95				56	0.97, 0.97	100, 100	0 95, 0.95	98.4, 98.4	0.96, 0.96	98.5, 98.4
4 97				84	0.99, 0.99	100, 100	0 96, 0.96	98.4, 98.5	0.97, 0.97	98.5, 98.5
4 96	citrate	1 0	-20	0	1.00, 1.00	100, 100	0 98, 0 98	98.8, 98.8	0.98, 0.98	98.1, 98.1
4 96				28	0.99, 1.00	100, 100	0 97, 0 97	98.3, 98.2	0.98, 0.98	98.5, 98 4
4 94				56	0.98, 0.97	100, 100	0.95, 0.95	98.3, 98.3	0 97, 0.96	98.6, 98 5
4 96				84	0.99, 0.99	100, 100	0.96, 0.96	98.4, 98.3	0.97, 0.96	98.5, 98.6

Example 2.

I. Analytical Methods

A. Reversed Phase (RP), Ion Exchange (IE) and Size Exclusion (SEC) Assays were conducted as described in Example 1.

II. Buffer Composition

All LIF samples were prepared by dilution of stock LIF solution containing 3.67 mg/ml LIF in 2 mM phosphate buffer, pH 6.42 to give the desired final LIF concentration (either 0.4 or 1.0 mg/ml) and composition of buffer components. The final composition of each solution contained 10 mM citrate buffer, 5% w/v sorbitol and 0.01% w/v Polysorbate 80. Samples differed in the final concentration of phosphate buffer (present from the original stock LIF solution) depending on the dilution factor. The 0.4 mg/ml LIF solutions contained 0.22 mM residual phosphate while the 1.0 mg/ml LIF solutions contained 0.54 mM residual phosphate. The composition of each buffer was as follows:

A. Citrate Buffer for 0.4 mg/ml LIF Formulations

Solution A: 11.22 mM sodium citrate dihydrate (Merck #1.06448)
5.61% w/v sorbitol (Sigma Chemicals #S1876)
0.0112% Polysorbate 80 (Sigma Chemicals #P1754)

Solution B: 11.22 mM citric acid monohydrate (Merck #1.00244)
5.61% w/v sorbitol (Sigma Chemicals #S1876)
0.0112% Polysorbate 80 (Sigma Chemicals #P1754)

Solutions A and B were mixed to give a final pH of 5.5. Formulations were prepared by combining 0.109 parts stock LIF solution and 0.891 parts buffer to give a final LIF concentration of 0.4 mg/ml, a final buffer concentration of 10 mM, a final sorbitol concentration of 5% w/v and a final Polysorbate 80 concentration of 0.01% w/v. The

measured osmolality of the final 0.4 mg/ml LIF formulation was 317 mOsm/kg.

B. Citrate Buffer for 1.0 mg/ml LIF Formulations

Solution A: 13.75 mM sodium citrate (Merck #1.06448)
6.88% w/v sorbitol (Sigma Chemicals #S1876)
0.0138% w/v Polysorbate 80 (Sigma Chemicals #P1754)

Solution B: 13.75 mM citric acid (Merck #1.00244)
6.88% w/v sorbitol (Sigma Chemicals S1876)
0.0138% w/v Polysorbate 80 (Sigma Chemicals P1754)

Solutions A and B were mixed to give a final pH of 5.5. Formulations were prepared by combining 0.272 parts stock LIF solution and 0.728 parts buffer to give a final LIF concentration of 1.0 mg/ml, a final buffer concentration of 10 mM, a final sorbitol concentration of 5% w/v and a final Polysorbate 80 concentration of 0.01% w/v. The measured osmolality of the final 1.0 mg/ml LIF formulation was 322 mOsm/kg.

II. Long Term Stability at 8°C and 25°C

A. Preparation of Samples for Storage at 8°C and 25°C

LIF formulations were prepared by dilution of stock LIF (3.67 mg/ml in 2 mM phosphate buffer, pH 6.42) with citrate buffer containing sorbitol and polysorbate 80 to give a final LIF concentration of 0.4 mg/ml or 1.0 mg/ml, a final buffer concentration of 10 mM, a final sorbitol concentration of 5% w/v and a final polysorbate 80 concentration of 0.01% w/v (see Section II). The theoretical pH was 5.5 and the actual pH of each sample was measured and recorded.

Under aseptic conditions in a laminar flow cabinet, the formulations were sterile filtered using 0.22 m Millex GV (Millipore) filters. The first 1.15 ml of each filtrate was set aside and the vial marked accordingly. The remaining volume was filtered into a sterile 50 ml polypropylene

tube. Aliquots of each formulation (1.15 ml/vial) were transferred using a multiple dispensing Eppendorf pipette with sterile tips into heat sterilised 2 ml glass vials and capped with sterile teflon lined rubber caps which were then crimped. Vials were labelled and duplicate vials of each formulation were retained for the initial analysis. The remaining vials were stored at either 8°C or 25°C.

B. Sample Analysis

All LIF samples were analysed undiluted along with standards according to the methods described in Example 1. At each time point, 2 vials were withdrawn from the incubators and approximately 200 µl was removed from each using a sterile 1 ml syringe and a sterile needle. These aliquots were placed into polypropylene autosampler vials and sealed with caps containing self-sealing septa to allow repeat injections from the same vial without evaporation. The original glass sample vials were then marked with the time point and placed at -80°C for repeat analysis (if required) or use in other studies.

Autosampler vials were transferred to the autosampler where they were maintained at 4°C throughout the three analytical runs. The same sample and standard autosampler vials were used for each of the three analyses with the RP (10 µl injection volume) being conducted first, followed by the IEC (100 µl injection volume) and then the SEC (10 µl injection volume). The complete RP run took approximately 20 hours, and the IEC and SEC runs took approximately 15 hours each. It was assumed that any further degradation over this storage time in the autosampler would be minimal (standard solutions at pH 6.42 stored under the same conditions showed no change over the complete analytical period).

Selected samples were also analysed for particulates using a Malvern Instruments Zetasizer 3000 particle size instrument. Samples were withdrawn from the storage vials using a syringe and placed in the sample cuvette. Samples were counted for 120 sec using a 200 m pinhole (to obtain the maximum signal), 90° scattering angle, and scattering source at 633 nm using a 10 mW He-Ne ion laser.

IV. Results

Data pertaining to solution pH, LIF concentration in mg/ml (determined by comparison to LIF standard solutions), and the area % for the main peak relative to the total peak area for all LIF related peaks in the chromatogram analysed using the three chromatographic methods are shown in Tables 18 and 19. For each set of samples, there was a slight decrease in solution pH of approximately 0.1 unit over the 92 day storage period.

1. Ion Exchange

A single main product (eluting at approximately 9 min) was seen in all samples stored at 8 and 25°. There was evidence of several minor degradation products in the ion exchange chromatograms, however, due to inadequate resolution between the different products, the exact number of products could not be determined. Samples prepared at pH 5.0 (initial study) and those at pH 5.5 (this study) stored at 8°C and 25°C for 8 weeks were compared. The chromatograms were normalised with respect to the retention time for the main peak to take into account slight changes in the chromatography between the two studies. In each case, the product distribution was similar with a higher proportion of the main degradation product noted in the pH 5.5 samples relative to the pH 5.0 samples.

The IEC results for the samples, wherein the main LIF peak was plotted as a percentage of the total area for all LIF related peaks in the chromatogram as a function of storage time illustrated the dependence of LIF stability on temperature. The relative stability under each storage condition was similar for the 0.4 and 1.0 mg/ml formulations. After 92 days storage at 8°C, 95-96% of the total peak area was present as the main LIF peak. Following storage at 25°C for 92 days, this value was reduced to approximately 56-58%.

The IEC stability data (main peak area expressed as a percentage of the total) obtained for samples at pH 5.5 with that from the previous study with samples prepared at pH 5.0 were compared. At 25°C, a slight increase in the rate of degradation was evident at pH 5.5.

2. Reversed Phase

RP chromatograms for the samples all displayed essentially the same elution characteristics. In all cases, the chromatograms showed the presence of only one main peak eluting at approximately 36 min.

The RP results wherein the measured concentration was plotted as a function of storage time illustrated the absence of significant change in the measured concentration over the storage period.

3. Size Exclusion

SEC chromatograms for the samples displayed essentially the same elution characteristics. In all cases, the chromatograms showed the presence of one main peak eluting at approximately 25 min and a minor peak eluting at approximately 21 min.

The SEC results wherein the measured concentration was plotted as a function of storage time illustrated the absence of significant change in the measured concentration over the storage period. Using the SEC method, there was no evidence of chain cleavage or crosslinking under the storage conditions studied.

4. Particle Size Analysis

Samples stored for 102 days at 8 and 25°C were analysed for particulates using a laser light scattering instrument. All of the samples analysed displayed a count rate of "0-0.5 kCps" which effectively means that the samples contained no particulates (i.e. no signal was measurable).

V. Summary

These studies demonstrated that formulations prepared at pH 5.5 were stable for up to 13

weeks when stored at 8°C with loss of the parent compound being approximately 3% as shown by IEC. After storage for 56 days at 8°C, the loss of LIF was approximately 2% in comparison to approximately 1% for pH 5.0 samples stored under the same conditions (data from the initial study). At 25°C, the rate of degradation at pH 5.5 was significantly increased with approximately 12% loss occurring in 4 weeks. In comparison, pH 5.0 samples showed a decrease in LIF concentration of approximately 7-9% after 4 weeks at 25°C. As in the initial study, no loss of LIF was detected by RP or SEC under any of the conditions studied.

Table 18 Summary of AM424 Stability for 0.4 mg/ml Formulations at pH 5.5 Following Storage at 8°C and 25°C

Measured pH	Buffer	Nominal LIF Conc. (mg/ml)	Storage Temp. (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc. (mg/ml)	SEC - Main Peak (area %)
5.61	citrate	0.4	8	0	0.42, 0.44	100, 100	0.40, 0.40	98.6, 98.6	0.40, 0.40	98.7, 98.7
5.56				14	0.38, 0.38	100, 100	0.37, 0.37	97.9, 98.0	0.39, 0.39	98.7, 98.7
5.59				29	0.41, 0.41	100, 100	0.39, 0.39	98.4, 98.3	0.41, 0.41	98.8, 98.7
5.51				42	0.40, 0.41	100, 100	0.37, 0.38	98.1, 97.8	0.40, 0.39	98.6, 98.7
5.47				56	0.39, 0.39	100, 100	0.39, 0.39	97.3, 97.1	0.40, 0.40	98.6, 98.7
5.48				77	0.39, 0.40	100, 100	0.38, 0.38	96.2, 96.3	0.39, 0.39	98.9, 98.9
5.48				92	0.42, 0.40	100, 100	0.37, 0.37	95.7, 95.8	0.38, 0.38	98.6, 98.7
5.61	citrate	0.4	25	0	0.42, 0.44	100, 100	0.40, 0.40	98.6, 98.6	0.40, 0.40	98.7, 98.7
5.57				14	0.38, 0.39	100, 100	0.35, 0.35	92.8, 92.8	0.39, 0.39	98.8, 98.9
5.59				29	0.41, 0.42	100, 100	0.35, 0.35	86.9, 86.8	0.41, 0.41	98.9, 99.0
5.52				42	0.41, 0.41	100, 100	0.31, 0.32	81.0, 81.8	0.38, 0.40	98.8, 99.0
5.48				56	0.39, 0.39	100, 100	0.29, 0.29	71.6, 71.9	0.41, 0.40	99.2, 99.0
5.48				77	0.41, 0.40	100, 100	0.26, 0.26	63.8, 64.0	0.40, 0.39	99.3, 99.1
5.48				92	0.40, 0.42	100, 100	0.23, 0.23	57.4, 57.7	0.39, 0.40	98.9, 99.2

Table 19 Summary of AM424 Stability for 1.0 mg/ml Formulations at pH 5.5 Following Storage at 8°C and 25°C

Measured pH	Buffer	Nominal LIF Conc. (mg/ml)	Storage Temp. (C)	Storage Time (days)	RP - Measured Conc. (mg/ml)	RP - Main Peak (area %)	IEC - Measured Conc. (mg/ml)	IEC - Main Peak (area %)	SEC - Measured Conc (mg/ml)	SEC - Main Peak (area %)
5.61	citrate	1.0	8	0	1.09, 1.08	100, 100	1.00, 1.00	98.6, 98.6	1.01, 1.01	98.6, 98.6
5.58				14	0.98, 0.99	100, 100	0.96, 0.96	97.7, 97.7	0.99, 0.99	98.6, 98.7
5.61				29	1.01, 1.02	100, 100	0.99, 0.99	97.5, 97.6	1.01, 1.01	98.5, 98.5
5.57				42	1.00, 1.01	100, 100	0.97, 0.97	97.3, 97.2	0.99, 0.98	98.4, 98.6
5.54				56	1.00, 0.99	100, 100	0.96, 0.96	96.8, 96.6	1.02, 1.02	98.4, 98.5
5.52				77	1.03, 1.02	100, 100	0.94, 0.94	96.0, 95.9	0.98, 0.99	98.5, 98.5
5.52				92	1.06, 1.04	100, 100	0.95, 0.94	95.3, 95.3	0.98, 0.98	98.4, 98.4
5.61	citrate	1.0	25	0	1.09, 1.08	100, 100	1.00, 1.00	98.6, 98.6	1.01, 1.01	98.6, 98.6
5.58				14	0.98, 0.98	100, 100	0.90, 0.90	91.7, 91.8	0.99, 0.99	98.7, 98.7
5.62				29	1.02, 1.01	100, 100	0.87, 0.87	85.6, 85.7	1.02, 1.02	98.7, 98.7
5.59				42	1.02, 1.01	100, 100	0.80, 0.80	80.0, 79.8	0.98, 0.98	98.8, 98.8
5.54				56	0.99, 1.02	100, 100	0.71, 0.71	68.9, 69.2	1.02, 1.03	98.8, 98.7
5.53				77	1.02, 1.03	100, 100	0.64, 0.64	61.0, 61.6	0.98, 0.98	98.9, 98.8
5.52				92	1.04, 1.09	100, 100	0.59, 0.58	56.0, 56.1	0.99, 0.99	98.8, 98.7

Example 3.

I. Sample Preparation

8°C and 25°C LIF Samples

LIF formulations were prepared by a dilution of stock LIF (3.67 mg/ml in 2 mM phosphate buffer) with citrate buffer containing sorbitol or NaCl to give a final LIF concentration of 0.05 or 0.4 mg/ml, a final buffer concentration of 10 mM, a final sorbitol concentration of 5% w/v or a final NaCl concentration of 0.9% w/v. The theoretical pH was 5.0 in all cases. Formulations were prepared and filled into vials as described previously.

II. Analytical Methods

Samples and standards were prepared as previously described. Analyses were conducted by RP and SEC and IEC was conducted using the Polycat A column.

The RP and SEC assays were the same as those described in Example 1. The IEC assay was conducted using a PolyLC PolyCAT A cation exchange, pH 6 phosphate buffer and a salt gradient. Detection was at 215 nm.

III. Results

Ion Exchange

IEC data for 0.4 mg/ml formulations are shown in Tables 20 and 21. The results were plotted with the main peak expressed as % of the initial since the % of total area values differ for the Pharmacia and Polycat A columns, and showed that at 25°C, the most stable formulations were the pH 5.0 citrate buffer containing sorbitol and Tween 80 and the pH 5.0 citrate containing NaCl. The least stable was the pH 5 citrate buffer containing only sorbitol and pH 5.5 citrate containing sorbitol and Tween 80 was somewhere in the middle.

SEC

SEC data for 0.05 and 0.4 mg/ml formulations are plotted with the main peak expressed as a % of the total area. There was some variability in the 0.05 mg/ml samples most likely due to the low concentration. There were no real trends for either buffer at 8°C or 25°C.

Freeze-Thaw Cycling

Freeze-thaw cycling studies for pH 5 citrate buffers containing sorbitol or NaCl were analysed by SEC. After the 5th cycle there was a trend toward a decrease in the main peak as a % of the total area and a slight increase in the pre-eluting high molecular weight peak.

Table 20 AM424 0.4 mg/ml Stability Following Storage at 8°C Measured by IEC

Storage Time (weeks)	Citrate/Sorbitol /Tween pH 5.0 Measured Conc. (mg/ml)[a]	Citrate/Sorbitol /Tween pH 5.0 Main Peak (area%)[a]	Citrate/Sorbitol /Tween pH 5.5 Measured Conc. (mg/ml)[a]	Citrate/Sorbitol /Tween pH 5.5 Main Peak (area%)[a]	Citrate/Sorbitol pH 5.0 Measured Conc. (mg/ml)[b]	Citrate/Sorbitol pH 5.0 Main Peak (area%)[b]	Citrate/NaCl pH 5.0 Measured Conc. (mg/ml)[b]	Citrate/NaCl pH 5.0 Main Peak (area %)[b]
0	0.36, 0.36	98.8, 98.7	0.40, 0.40	98.6, 98.6	0.28, 0.28	73.1, 73.2	0.28, 0.28	72.9, 72.8
2	0.38, 0.38	98.4, 98.4	0.37, 0.37	97.9, 98.0	0.27, 0.28	72.1, 72.0	0.27, 0.28	71.9, 72.4
4	0.38, 0.37	98.4, 98.5	0.39, 0.39	98.4, 98.3	0.29, 0.28	73.3, 72.8	0.28, 0.28	74.1, 73.9
6	0.38, 0.37	97.8, 97.9	0.37, 0.38	98.1, 97.8	0.30, 0.29	73.4, 72.4	0.30, 0.30	73.9, 73.4
8	0.37, 0.37	97.7, 97.5	0.39, 0.39	97.3, 97.1	0.29, 0.29	71.9, 71.7	0.28, 0.29	71.6, 72.3

[a] Pharmacia Mono S Column
[b] PolyCAT A Column

Table 21 AM424 0.4 mg/ml Stability Following Storage at 25°C Measured by IEC

Storage Time (weeks)	Citrate/Sorbitol /Tween pH 5.0 Measured Conc. (mg/ml)[a]	Citrate/Sorbitol /Tween pH 5.0 Main Peak (area%)[a]	Citrate/Sorbitol /Tween pH 5.5 Measured Conc. (mg/ml)[a]	Citrate/Sorbitol /Tween pH 5.5 Main Peak (area%)[a]	Citrate/Sorbitol pH 5.0 Measured Conc. (mg/ml)[b]	Citrate/Sorbitol pH 5.0 Main Peak (area%)[b]	Citrate/NaCl pH 5.0 Measured Conc. (mg/ml)[b]	Citrate/NaCl pH 5.0 Main Peak (area %)[b]
0	0.36, 0.36	98.8, 98.7	0.40, 0.40	98.6, 98.6	0.28, 0.28	73.1, 73.2	0.28, 0.28	72.9, 72.8
2	0.36, 0.37	95.4, 95.1	0.35, 0.35	92.8, 92.8	0.26, 0.26	64.5, 67.9	0.27, 0.25	69.7, 67.4
4	0.34, 0.34	92.0, 91.8	0.35, 0.35	86.9, 86.8	0.24, 0.25	61.3, 61.6	0.26, 0.25	67.3, 66.4
6	0.33, 0.34	87.0, 87.4	0.31, 0.32	81.0, 81.8	0.24, 0.24	58.9, 59.0	0.26, 0.26	62.7, 62.9
8	0.30, 0.30	77.8, 77.9	0.29, 0.29	71.6, 71.9	0.21, 0.20	54.0, 51.7	0.24, 0.24	59.5, 59.2

[a] Pharmacia Mono S Column
[b] PolyCAT A Column

Example 4.

Preferred compositions comprise:

- LIF in a concentration of 400 to 1000 mg/ml
- pH of about 4.0 - 6.0
- surfactant
- isotonicity agent
- buffer.

Particularly preferred compositions are those wherein the pH range is about 4.5 - 5.5.

Example 5.

A particularly preferred composition comprises:

- LIF in a concentration of 400 to 1000 mg/ml
- pH of about 5.0
- 5% w/w sorbitol
- 0.01% polysorbate 80
- citrate or acetate buffer.

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and compounds referred to or indicated in this specification, individually or collectively, and any and all combinations of any two or more of said steps or features.

CLAIMS

1. A composition comprising Leukaemia Inhibitory Factor (LIF) or a derivative or homologue thereof and a stabilizing agent facilitating chemical and/or physical stability of LIF in the composition and one or more pharmaceutically acceptable carriers and/or diluents.

2. A composition according to claim 1 wherein the stabilizing agent facilitates reduced aggregation of LIF.

3. A composition according to claim 1 or 2 wherein the stabilizing agent facilitates a reduction in the deamidation of LIF.

4. A composition according to claim 1 or 2 or 3 wherein the pH of the composition is from between about 3.5 and 6.5.

5. A composition according to claim 3 wherein the pH of the composition is from between about 3.5 and 6.5.

6. A composition according to claim 1 or 5 wherein the stabilizing agent is an isotonicity agent, an agent which increases or maintains the conformational stability of LIF or its derivatives or homologues or a surfactant or functional equivalents thereof.

7. A composition according to claim 6 wherein the stabilizing agent is an isotonicity agent selected from a polyhydric alcohol, a pharmaceutically acceptable salt, a buffer species, a sugar and a pharmaceutically acceptable polymeric compound.

8. A composition according to claim 7 wherein the polyhydric alcohol is sorbitol.

9. A composition according to claim 6 wherein the surfactant is an anionic, cationic, amphoteric or non-ionic surfactant.

10. A composition according to claim 9 wherein the surfactant is selected from a fatty alcohol, a glyceryl ester and a fatty acid ester of a fatty alcohol or other alcohol.

11. A composition according to claim 6 wherein the stabilizing agent is selected from a polysorbate, a polyoxyethylene derivative and a pharmaceutically acceptable polyoxyethylene-polyoxypropylene copolymer.

12. A composition according to claim 7 wherein the buffer species is selected from a phosphate, citrate and acetate buffer.

13. A composition according to claim 12 wherein the buffer species is a citrate or acetate buffer.

14. A composition comprising Leukaemia Inhibitory Factor (LIF) and one or more pharmaceutically acceptable carriers and/or diluents and wherein the composition has a pH of between 3.5 and 6.5.

15. A composition according to claim 6 wherein the aggregation of LIF over time is reduced.

16. A composition according to claim 6 or 7 wherein the deamidation of LIF over time is reduced.

17. A composition according to claim 14 where the pH is maintained by the presence of a buffer species selection from a phosphate, citrate and acetate buffer.

18. A composition according to claim 17 wherein the buffer species is a citrate or acetate buffer.

19. A composition according to claim 14, 17 or 18 wherein the pH is between from about 4.5 and about 5.5.

20. A composition according to claim 1 or 14 wherein LIF is present in an amount from about 0.1 µg/ml to about 100 mg/ml.

21. A method for preparing a composition comprising Leukaemia Inhibitory Factor (LIF) or a derivative or homologue thereof and which exhibits reduced deamidation and/or agglutination of LIF or its derivative or homologues over time said method comprising admixing LIF or its derivative or homologue with a stabilizing agent.

22. A method according to claim 21 wherein the stabilizing agent is a isotonicity agent, an agent which increases or maintains the conformational stability of LIF or its derivatives or homologues or a surfactant or functional equivalents thereof.

23. A method according to claim 22 wherein the stabilizing agent is an isotonicity agent selected form a polyhydric alcohol, a pharmaceutically acceptable salt, a buffer species, a sugar and a pharmaceutically acceptable polymeric compound.

24. A method according to claim 23 wherein the polyhydric alcohol is sorbitol.

25. A method according to claim 22 wherein the surfactant is an anionic, cationic, amphoteric or non-ionic surfactant.

26. A method according to claim 25 wherein the surfactant is selected from a fatty alcohol, glyceryl ester and a fatty acid ester of a fatty alcohol or other alcohol.

27. A method according to claim 22 wherein the stabilizing agent is selected from a polysorbate, a polyoxyethylene derivative and a pharmaceutically acceptable polyoxyethylene-polyoxypropylene copolymer.

28. A method according to claim 23 wherein the buffer species is selected from a phosphate, citrate and acetate buffer.

29. A method according to claim 28 wherein the buffer species is a citrate or acetate buffer.

30. A method according to any of claims 22 to 29 further comprising adjusting the pH to between from about 3.5 and about 6.5.

31. A method according to claim 30 wherein the pH is between from about 4.5 and about 5.5.

32. A method according to any one of the claims 22 to 31 further comprising admixing one or more pharmaceutically acceptable carriers and/or diluents.

33. Use of a stabilizing agent in the manufacture of a composition exhibiting improved chemical and/or physical stability of Leukaemia Inhibitory Factor (LIF) or a derivative or homologue thereof.

34. Use according to claim 33 wherein the stabilizing agent is an isotonicity agent selected from a polyhydric alcohol, a pharmaceutically acceptable salt, a buffer species, a sugar and a pharmaceutically acceptable polymeric compound.

35. Use according to claim 34 wherein the polyhydric alcohol is sorbitol.

36. Use according to claim 34 wherein the surfactant is an anionic, cationic, amphoteric

or non-ionic surfactant.

37. Use according to claim 36 wherein the surfactant is selected from a fatty alcohol, glyceryl ester and a fatty acid ester of a fatty alcohol or other alcohol.

38. Use according to claim 33 where the stabilizing agent is selected from a polysorbate, a polyoxyethylene derivative or a pharmaceutically acceptable polyoxyethylene-polyoxypropylene copolymer.

39. Use according to claim 34 wherein the buffer species is selected from a phosphate, citrate and acetate buffer.

40. Use according to claim 39 wherein the buffer species isa a citrate or acetate buffer.

41. Use according to any one of claims 33 to 40 where the pH of the composition is between from about 3.5 to about 6.5.

42. Use according to any one of claims 41 wherein the pH is between from about 4.5 and about 5.5.

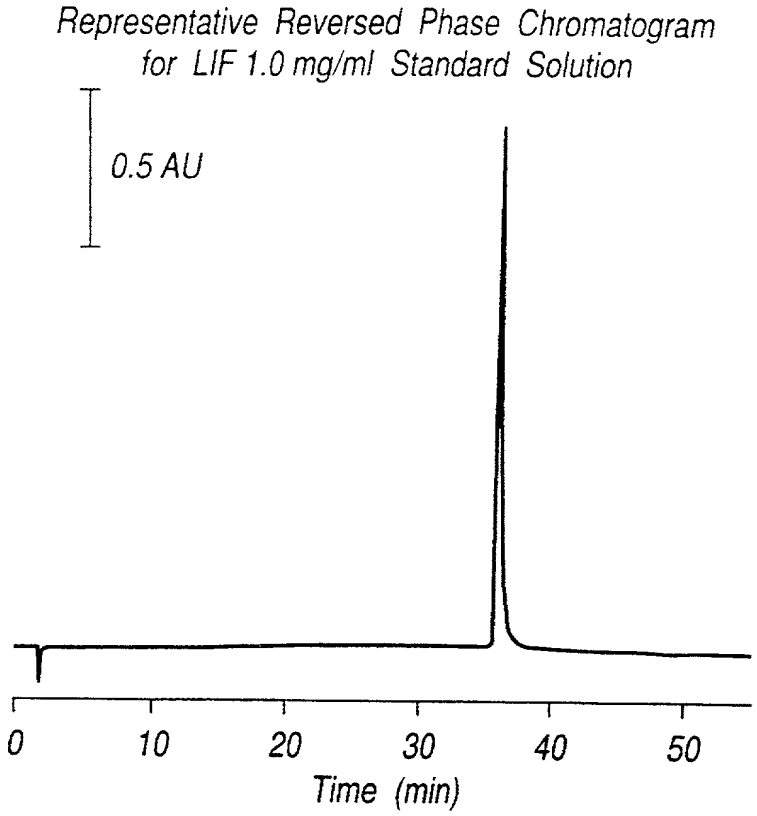

Representative Reversed Phase Chromatogram
for LIF 1.0 mg/ml Standard Solution
0.5 AU
0
10
20
30
40
50
Time (min)

Fig.1

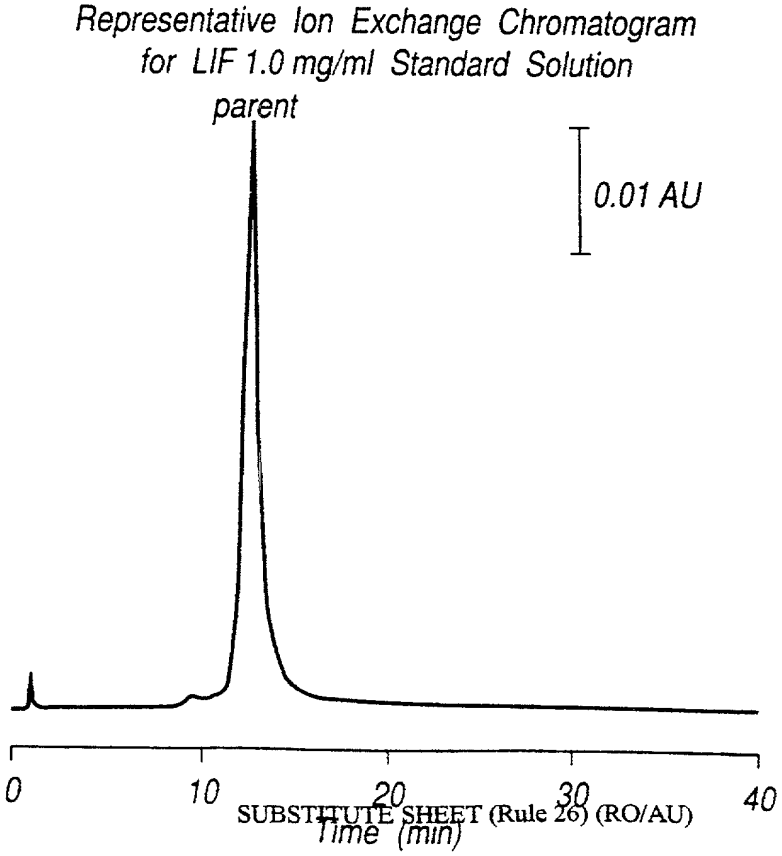

Representative Ion Exchange Chromatogram
for LIF 1.0 mg/ml Standard Solution
parent
0.01 AU
0
10
20
30
40
Time (min)

Fig.2

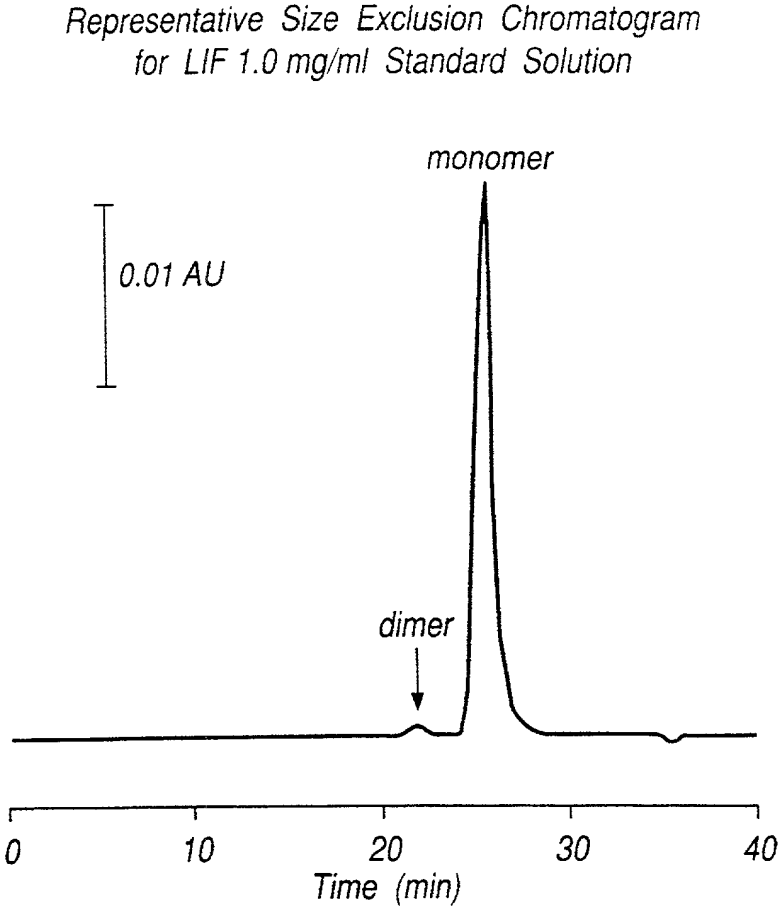

Representative Size Exclusion Chromatogram
for LIF 1.0 mg/ml Standard Solution
monomer
0.01 AU
dimer
0
10
20
30
40
Time (min)

Fig.3

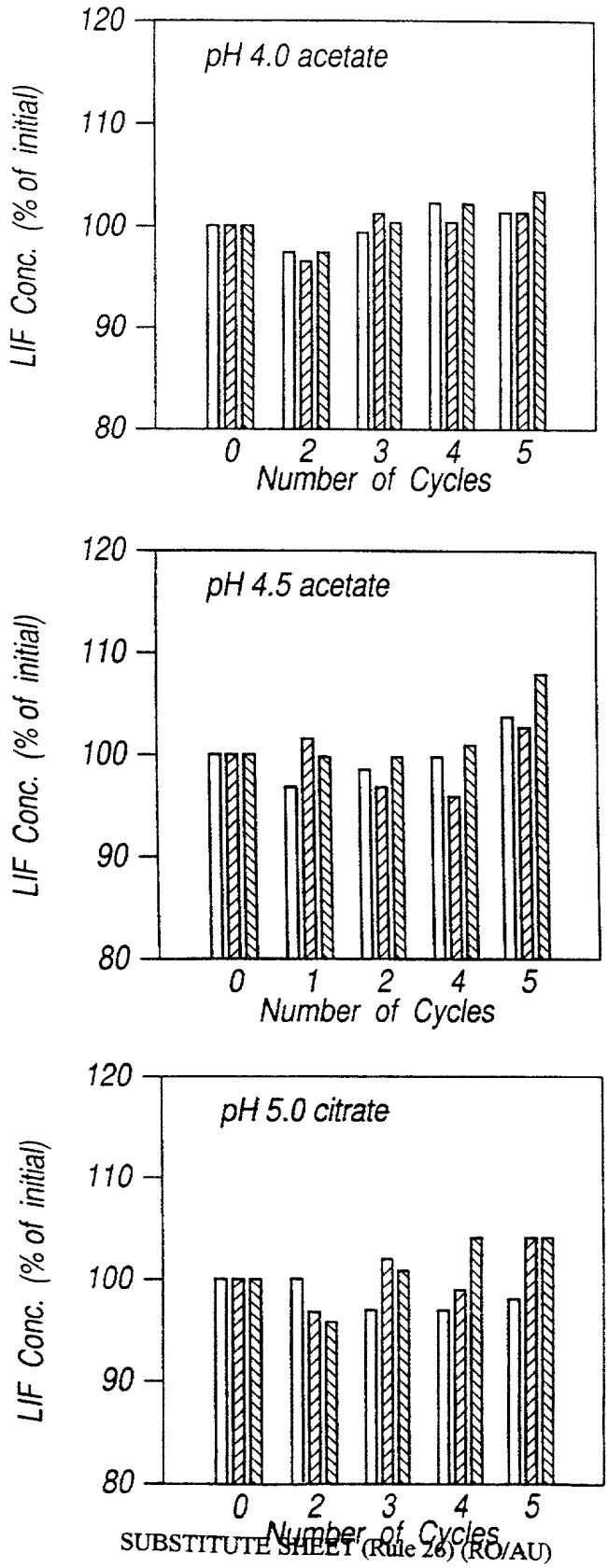

Individual Freeze/Thaw Cycling Results
pH 4.0 acetate
LIF Conc. (% of initial)
120
110
100
90
80
0 2 3 4 5
Number of Cycles
pH 4.5 acetate
LIF Conc. (% of initial)
120
110
100
90
80
0 1 2 4 5
Number of Cycles
pH 5.0 citrate
LIF Conc. (% of initial)
120
110
100
90
80
0 2 3 4 5
Number of Cycles

Fig.4

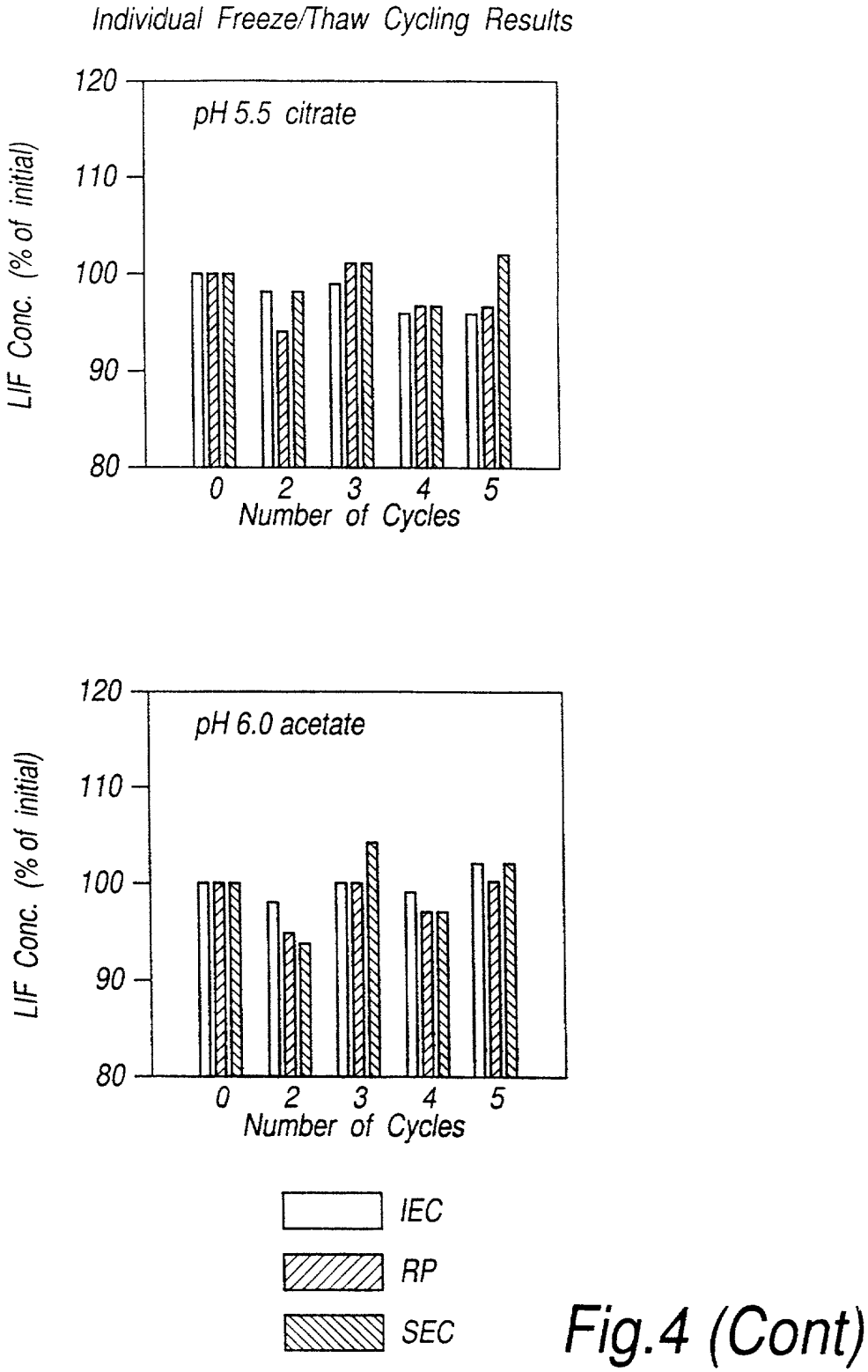

Individual Freeze/Thaw Cycling Results
pH 5.5 citrate
LIF Conc. (% of initial)
120
110
100
90
80
0
2
3
4
5
Number of Cycles
pH 6.0 acetate
LIF Conc. (% of initial)
120
110
100
90
80
0
2
3
4
5
Number of Cycles
IEC
RP
SEC

Fig.4 (Cont)

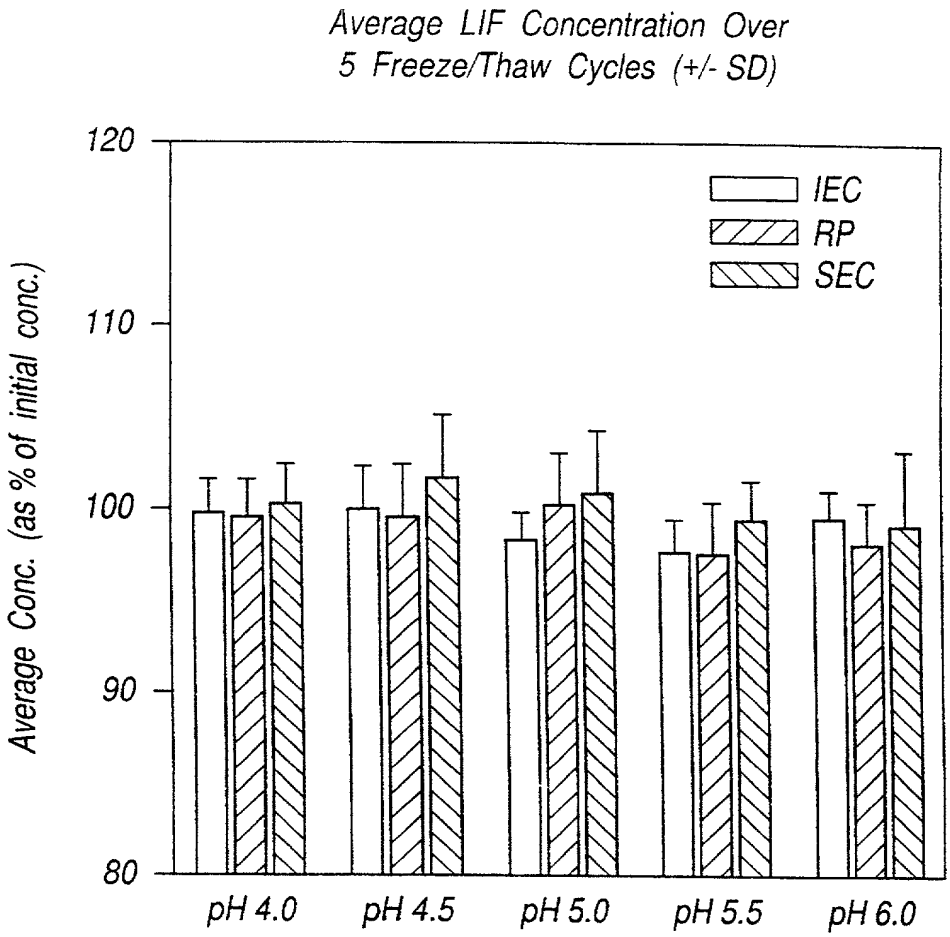

Average LIF Concentration Over
5 Freeze/Thaw Cycles (+/- SD)
Average Conc. (as % of initial conc.)
120
110
100
90
80
IEC
RP
SEC
pH 4.0
pH 4.5
pH 5.0
pH 5.5
pH 6.0

Fig.5

COMBINED DECLARATION FOR PATENT APPLICATION AND POWER OF ATTORNEY

Includes Reference to PCT International Applications

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

COMPOSITIONS OF LEUKAEMIA INHIBITORY FACTOR

the specification of which (check only one item below):

☐ is attached hereto.

☐ was filed as United States application

Serial No. _____

on _____,

and was amended

on _____ (if applicable).

☑ was filed as PCT international application

Number PCT/AU98/00981

on 26 November 1998,

and was amended under PCT Article 19

on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowlege the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

PRIOR FOREIGN/PCT APPLICATION(S) AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. 119:

COUNTRY (if PCT indicate PCT)	APPLICATION NUMBER	DATE OF FILING (day, month, year)	PRIORITY CLAIMED UNDER 35 USC 119
Australia	PP 0531	26 November 1997	☑ YES ☐ NO
			☐ YES ☐ NO
			☐ YES ☐ NO
			☐ YES ☐ NO
			☐ YES ☐ NO

Combined Declaration For Patent Application and Power of Attorney (Continued)
(Including Reference to PCT International Application)

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) or PCT international application(s) designating the United States of America that is are listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior application(s) in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowlege the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56(a) which occurred between the filing date of the prior application(s) and the national or PCT international filing date of this application:

PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S. FOR BENEFIT UNDER 35 U.S.C. 120:

U S APPLICATIONS		STATUS (Check one)		
U S APPLICATION NUMBER	U S FILING DATE	PATENTED	PENDING	ABANDONED

PCT APPLICATIONS DESIGNATING THE U.S.

PCT APPLICATION NO	PCT FILING DATE	U S SERIAL NUMBERS ASSIGNED (if any)			

OIPE JUL 21 2000 PATENT & TRADEMARK OFFICE

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. Stephan D. Murphy, Reg. No. 22,002; Leopold Presser, Reg. No. 19,827; William C. Roch, Reg. No. 24,972; Kenneth L. King, Reg. No. 24,223; Frank S. DiGiglio, Reg. No. 31,346; Paul J. Esatto, Jr., Reg. No. 30,749; John S. Sensny, Reg. No. 28,757; Mark J. Cohan, Reg. No. 32,211; Richard L. Catania, Reg. No. 32,608 and Donald T. Black, Reg. No. 27,999.

10

Send Correspondence to:	Direct Telephone Calls to: (name and telephone number)
Scully, Scott, Murphy & Presser 400 Garden City Plaza Garden City, NY 11530	Leopold Presser (516) 742-4343

		FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME
201	FULL NAME OF INVENTOR	CHARMAN	Susan	Ann
	RESIDENCE & CITIZENSHIP	CITY: Brighton AUX	STATE OR FOREIGN COUNTRY: Victoria, Australia	COUNTRY OF CITIZENSHIP: United States
	POST OFFICE ADDRESS	POST OFFICE ADDRESS: 55 Whyte Street	CITY: Brighton	STATE & ZIP CODE/COUNTRY: Victoria 3186 Australia
202	FULL NAME OF INVENTOR	RADFORD	Anthony	John
	RESIDENCE & CITIZENSHIP	CITY: Kew, AUX	STATE OR FOREIGN COUNTRY: Victoria, Australia	COUNTRY OF CITIZENSHIP: Australia
	POST OFFICE ADDRESS	POST OFFICE ADDRESS: 2 Pakington Street	CITY: Kew,	STATE & ZIP CODE/COUNTRY: Victoria3101 Australia
203	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	STATE & ZIP CODE/COUNTRY

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

SIGNATURE OF INVENTOR 201	SIGNATURE OF INVENTOR 202	SIGNATURE OF INVENTOR 203
× Susan A. Charman	× A. Radford	
DATE × 13 July 2000	DATE × 12 July 2000	DATE

[ ] Signature for fourth and subsequent joint inventors.
Number of pages added _____.

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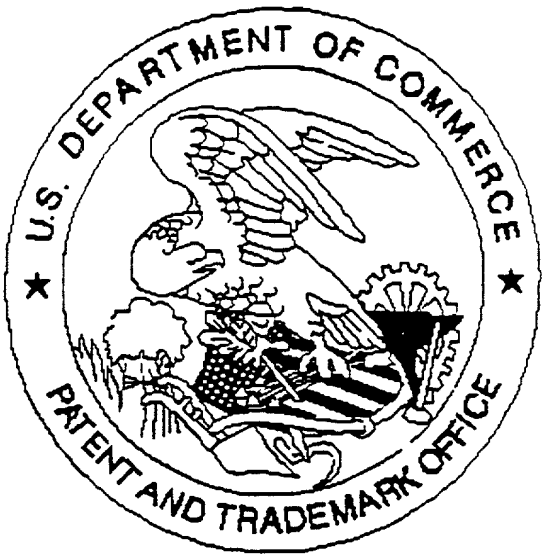

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